

A NEW APHID ON GUAYULE AND NOTES  
ON OTHER SPECIES OF CEROSIPHA<sup>1</sup>E. O. ESSIG<sup>2, 3</sup>

ON JULY 13, 1942, during an investigation of insects attacking guayule, *Parthenium argentatum*, in the United States government nurseries at Salinas, California, W. H. Lange discovered on some slightly wilted seedlings, a new root-infesting species of aphid. In preparing the description of this insect, comparisons had to be made with other known species of the genus *Cerosipha* to which it belongs. Since there is some confusion concerning the relatively few species relegated to this genus, enough descriptive and illustrative material has been included to make possible ready recognition of all the members.

## THE GUAYULE APHID

*Cerosipha californica* n. sp.

Lange found the aphids in small bark and root cavities made by wireworms and other insects. Frequently but a single specimen occupied such a cavity. A thorough search revealed relatively few aphids present and on young nursery stock only. So far no more have been found in the nurseries and none at all in the older field plantings. In all, only 1 alate and 4 apterous mature parthenogenetic females, 12 young, and 7 nymphs were collected on guayule.

In my collection there was also a single balsam slide mount containing 3 alates of a species taken on the leaves of French prune in the orchard of the University of California, Davis, on November 8, 1928. These alates were labeled *Cerosipha* sp. In order to study them better they were cleared and remounted in euparal, a single specimen to a slide. In all respects they appear to be identical with the single alate taken on guayule and are treated as paratypes of the species.

Additional specimens of this species were recently taken on the roots of potatoes at Bakersfield, California, on May 8, 1944, by Gordon L. Smith.

In checking over some mounted specimens collected by W. C. Look on tomato at the Waipaha School Gardens, Waipaha, Oahu, Territory of Hawaii, September 9, 1940, I find that three alate specimens provisionally determined as *Vesiculaphis caricis* (Full.) prove to be this species. An important variation in the wing venation is the frequent extension of the second fork of the media vein to almost the margin of the wings as in one specimen and its omission as

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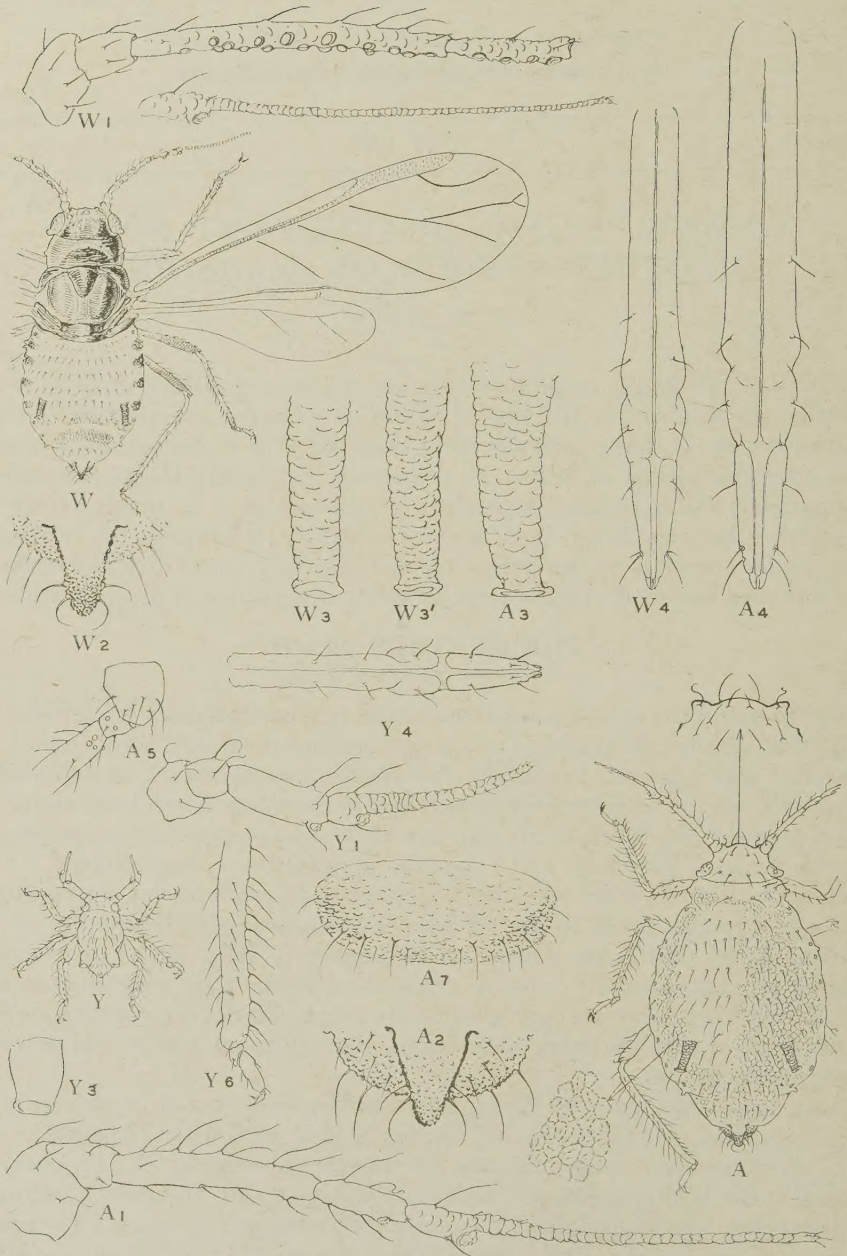


Fig. 1.—The guayule aphid, *Cerosipha californica* n. sp.: A, apterous viviparous females; W, winged or alate viviparous females; Y, young of first instar. 1, Antenna; 2, cauda; 3, cornicle (W3, on prune; W3', on guayule); 4, rostrum; 5, base of hind leg; 6, hind tibia and tarsus; 7, genital plate.



in the other two specimens. Variations of this sort are confusing and often lead to misidentification.

*Color*.—According to notes made by Lange, the living forms on the roots of guayule are dark grayish green, not greatly different from the general color of the bark and foliage of the host plant. Mounted specimens show the antennae, legs, cornicles, and cauda to be dusky. Other dark markings on the alates are shown in the accompanying drawings.

*Winged Viviparous Female* (fig. 1, *W*).—Length 1.8 mm; width 0.8 mm; length of antenna 1.05 mm (I, 0.08 mm; II, 0.05 mm; III, 0.33 mm; IV, 0.09 mm; V, 0.50 mm, the base being 0.10 mm and the filament 0.40 mm); length of forewing 2.50 mm. Body hairs rather fine and mostly curved. Hairs on the antennae may equal the width of the segments; those on the tibiae are stiffer and also nearly equal the width. A pair of small lateral tubercles are present on the prothorax; a double pair on abdominal segment II, and a single pair on segment VIII. Left member of antennal segment III with 18 and right member with 22 large circular secondary sensoria. These sensoria are distributed over most of the length in an irregular row. Left member of segment IV with 3 and right member with 2 large circular secondary sensoria. The permanent large sensoria on IV and V are conspicuously fringed with hairs. Rostrum somewhat blunt and with few short hairs, 0.48 mm long, and extending to the third coxae. Wings normal, with radial sector and second fork of media near tip. Cornicles dusky, cylindrical or slightly swollen with a constriction just before the well-developed distal flange, coarsely imbricated, 2.2 mm long. Cauda dusky, tapering gradually from the wide base to a blunt rounded tip, armed with 2 pairs of curved spines.

*Apterous Viviparous Female* (fig. 1, *A*).—Length of antenna 0.84 mm (I, 0.7 mm; II, 0.05 mm; III, 0.20 mm; IV, 0.09 mm; V, 0.43 mm, the base being 0.08 mm and the filament 0.35 mm). Body clothed with rather long fine spines or hairs, which are straight, slightly curved, or crooked, and of variable length, those on the tibiae and antennae mostly longer than the width of the segments. Epidermis of cleared specimens reticulated and densely beset with microtrichialike skin elevations. These latter are arranged in lines on dorsum and venter. A pair of simple lateral tubercles on the prothorax and on abdominal segments II and VIII. Antennae 5-segmented; I, noticeably large and almost as wide as long; secondary sensoria absent; a single large circular and fringed primary sensorium present on IV and on V; marginal sensoria were not noted on V, but there appear to be three rather prominent scalelike processes on the lower side of the large sensorium which may correspond to marginal sensoria. These are best seen in the young as illustrated. Rostrum extends to first abdominal segment. Cornicles similar to those of the alate, 0.25 mm long. Cauda, cone-shaped with wide base, 0.10 mm long, and with 2 pairs of curved spines. Genital plate dusky and as illustrated.

*Young, 1st instar*, is noticeably hairy or spiny, with 5-segmented antennae of which the first segment is conspicuously large. Rostrum extending to the middle of the abdomen, with slightly tapering cornicles. Legs stout and hairy.

This species has been compared with other members of the genus and relationships are indicated in the key and illustrations. The winged forms superficially resemble *Rhopalosiphum prunifoliae* (Fitch), but the 5-segmented antennae, the close proximity of the second fork of the media to the marginal tip of the wing, and the shape and spination of the cauda readily separate the two.

A single alate viviparous female collected on guayule has been designated as the type. The three alates taken on prune at Davis and the mature apterous viviparous females and young feeding on guayule are designated as paratypes in my collection. Paratypes, alate (on prune) and apterous, have been deposited in the collections of the United States Bureau of Entomology and Plant Quarantine, Washington, D. C.

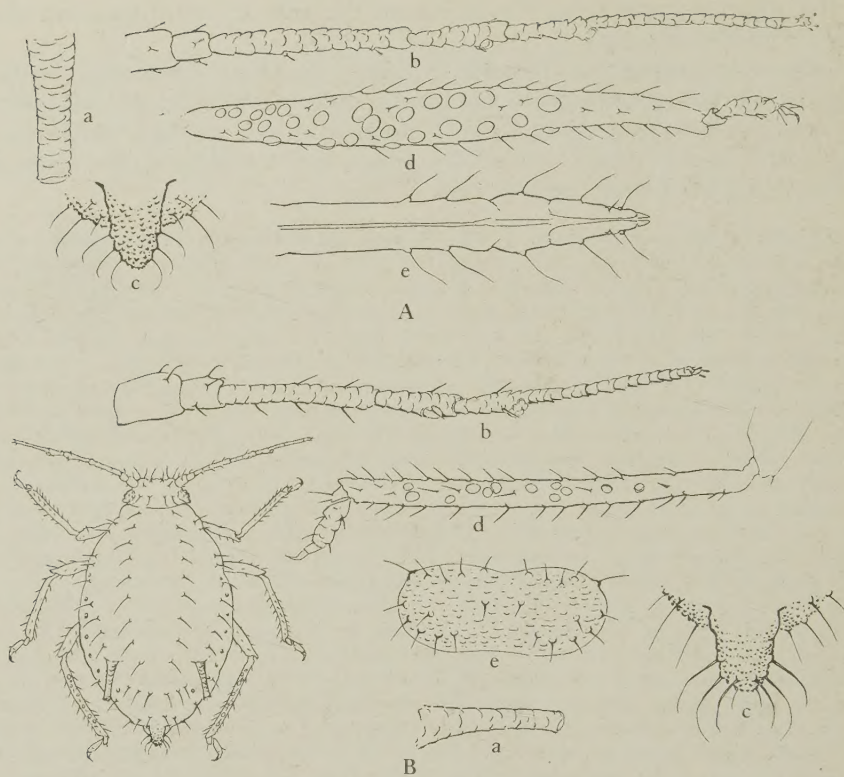


Fig. 2.—The raspberry aphid, *Cerosipha rubifolii* (Thomas): A, oviparous female from Ohio: a, cornicle; b, antenna; c, cauda; d, hind tibia; e, rostrum. B, oviparous female from California: a, cornicle; b, antenna; c, cauda; d, hind tibia; e, genital plate.

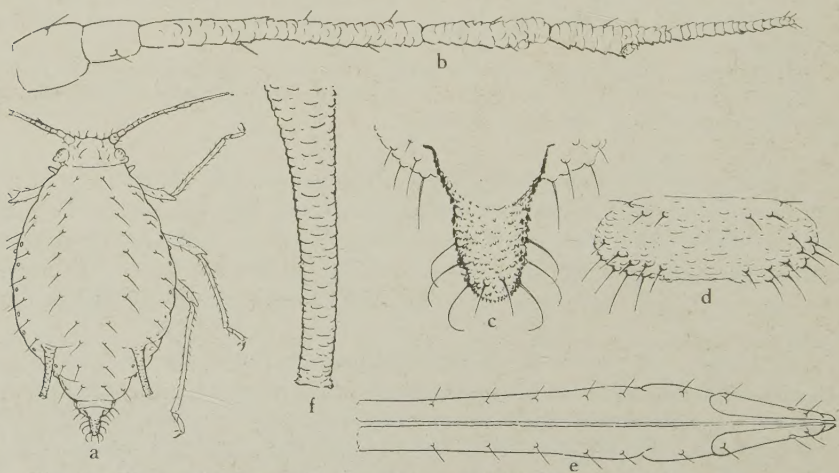


Fig. 3.—The raspberry aphid, *Cerosipha rubifolii* (Thomas) (?), apterous viviparous female from California: a, adult; b, antenna; c, cauda; d, genital plate; e, rostrum; f, cornicle.



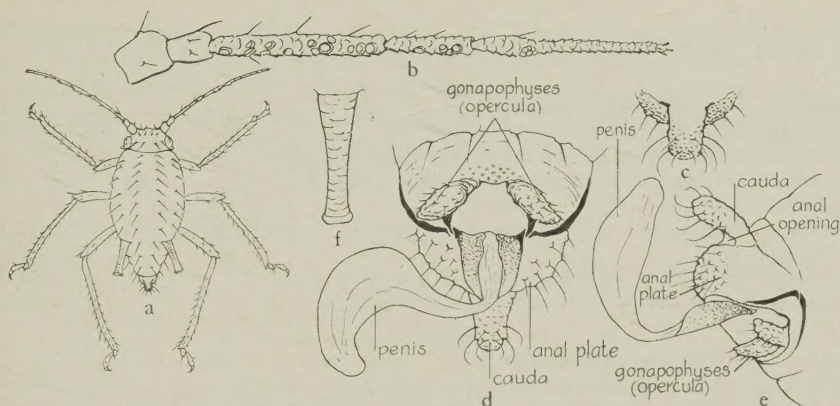


Fig. 4.—The raspberry aphid, *Cerosipha rubifolii* (Thomas) (♀), male from California: a, adult; b, antenna; c, cauda; d, posterior tip of abdomen (pygidium), ventral aspect; e, same, lateral aspect; f, cornicle.

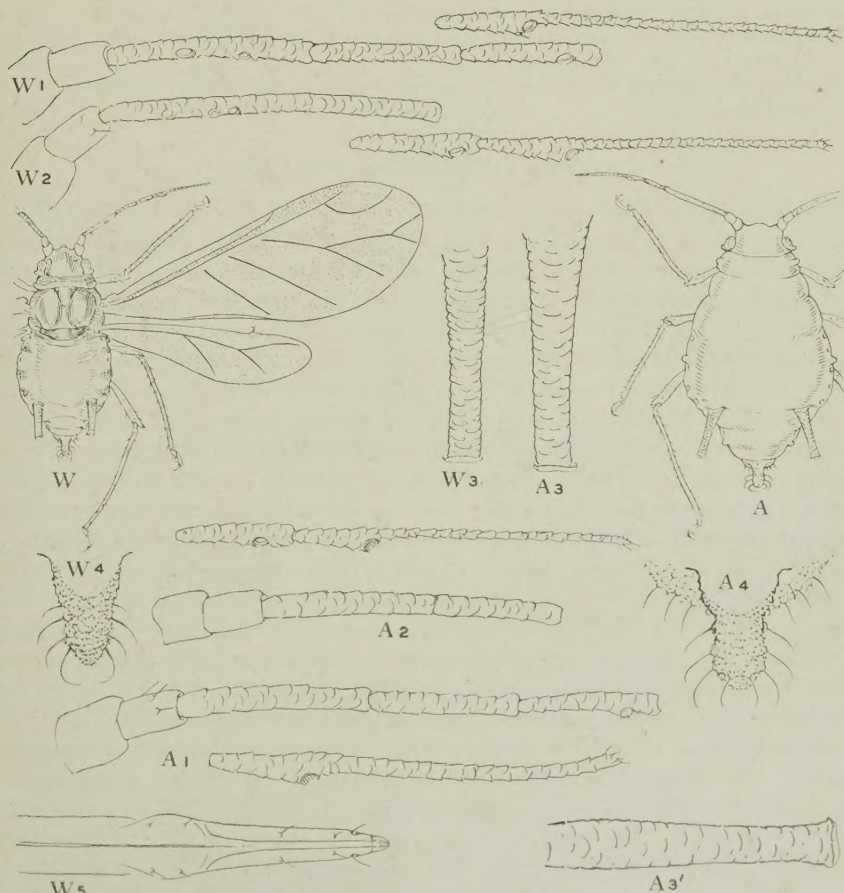


Fig. 5.—The strawberry root aphid, *Cerosipha forbesi* (Weed): A, apterous viviparous female; W, winged viviparous female; 1, 6-segmented antenna; 2, 5-segmented antenna; 3, cornicle; 3', cornicle from California apterous female; 4, cauda; 5, rostrum.

## KEY TO KNOWN SPECIES OF THE GENUS CEROSIPHA

## ALATES

1. Forewing with radial sector present..... 2  
Forewing with radial sector usually absent..... *araliae-radicis*
2. Secondary sensoria present only on antennal segment III..... 3  
Secondary sensoria on both antennal segments III and IV..... *californica*
3. Antennae rarely more than 5-segmented..... 4  
Antennae 5-segmented or 6-segmented; from 1 to 4 secondary sensoria on antennal segment III..... *forbesi*
4. Antennal segment III with 4 to 7 secondary sensoria..... *rubifolii*  
Antennal segment III with 10 to 19 secondary sensoria..... *roripae*

## APTERAE

1. Body hairs short and relatively few..... 2  
Body hairs long and prominent, especially on the legs..... *californica*
2. Filament of apical antennal segment at least five times the length of the base..... *araliae-radicis*  
Filament of apical antennal segment less than five times the length of the base..... 3
3. Antennal segment III without sensoria..... 4  
Antennal segment III normally with sensoria..... *roripae*
4. Rostrum very long and slender throughout, segment III only slightly swollen; cornicles about twice as long as the cauda..... *forbesi*  
Rostrum with segment III noticeably swollen; cornicles only slightly longer than the cauda..... *rubifolii*

## NOTES ON OTHER SPECIES

*The Raspberry Aphid*.—Named in the genus *Sipha* by Thomas in 1879, the raspberry aphid, *Cerosipha rubifolii* (Thomas) (figs. 2 to 4), appears to be a leaf-feeding species, which has been widely reported from North America. Judging from specimens examined, either the species is quite variable in structure, or there are two or even three species confused under the one name. For instance, the species occurring in California has unusually long cornicles; again, a considerable number of the specimens furnished by G. F. Knowlton from Oregon, Washington, and Utah are so extremely small and otherwise different from eastern material as to be questionable. Lacking an adequate amount of material of the various forms, I am unable to settle this matter now.

Alate viviparous females usually have 5-segmented antennae, and in a single apterous viviparous female one member was 5-segmented and the other 6-segmented. This species is much more consistent in having 5-segmented antennae than is *Cerosipha forbesi*. Secondary circular sensoria occur on segment III and vary from 3 to 7 in number; the average was 5 for 23 antennae examined.

Sexuals, especially the apterous females, are apparently rather common. These have numerous large, nearly circular sense-pits over the entire hind tibiae except the apical portions. These sensoria vary considerably in size, and in numbers from 13 to 26 or more. A single apterous male taken on raspberry at Palo Alto, California, November 1, 1928, by L. M. Smith, has 10 and 11 small circular sensoria on antennal segment III and 3 and 6 on segment IV.

*The Strawberry Root Aphid*.—Named in the genus *Aphis* and later transferred to the genus *Cerosipha*, the strawberry aphid, *Cerosipha forbesi* (Weed) (fig. 5), might appropriately be placed in either genus because of the division of the antennae into either 5 or 6 segments. An examination of 27



alates collected in California revealed that 7 had 6-segmented antennae, 13 had 5-segmented antennae, and 7 had one member 6-segmented and the other 5-segmented. In many of those with 5-segmented antennae there is a distinct distortion in the middle region of segment III indicating the place of division to form the extra segment, and in practically all cases segment III in the 5-

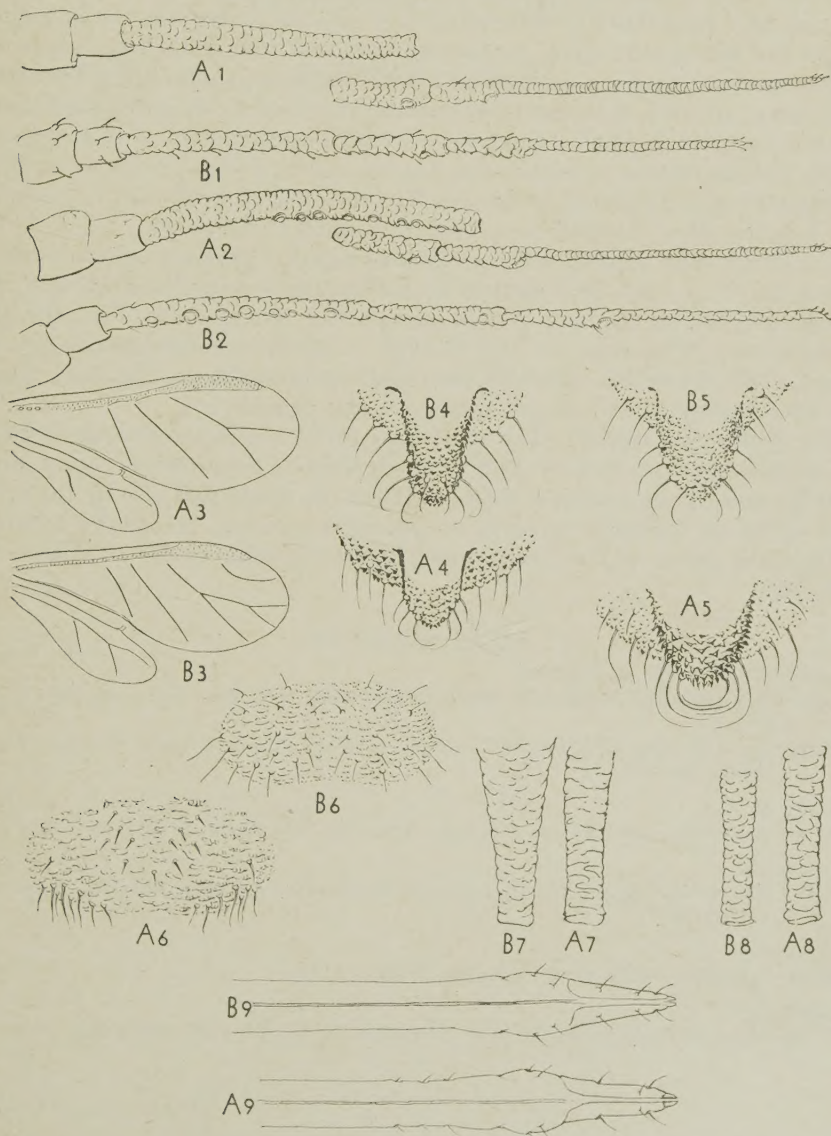


Fig. 6.—A, Structural parts of the aralia-root aphid, *Cerosipha araliae-radialis* Strom; B, same for the raspberry aphid, *C. rubifolii* (Thomas), from Illinois; 1, antenna of apterous viviparous female; 2, antenna of alate viviparous female; 3, wings; 4, cauda of apterae; 5, cauda of alates; 6, genital plates; 7, cornicles of apterae; 8, cornicles of alates; 9, rostrums.

segmented antennae equals or nearly equals in length the combined segments III and IV in the 6-segmented forms. Of 15 apterous adults examined all had 5-segmented antennae except one, which had both members 6-segmented. If the segment is not divided (5-segmented antenna) the sensoria are distributed over the basal half; if it is divided (6-segmented antenna) they occur over the whole shorter, basal segment as illustrated. Antennal segment III may have from 0 to 4 circular secondary sensoria which vary considerably in size but are usually equal in diameter to about half that of the segment.

The number of sensoria on segment III in 5-segmented antennae differ little, often not at all, from those with 6-segmented antennae. In 22 individuals there was but 1 without sensoria; there were 5 with 1 sensorium on both members; 7 with 1 sensorium on 1 member and 2 on the other; 2 with 3 sensoria on both members; 4 with 2 sensoria on one and 3 on the other; 2 with 3 on one member and 4 on the other; 1 with 5 on one member and 1 on the other.

*The Aralia-Root Aphid.*—The aralia-root aphid, *Cerosipha araliae-radialis* Strom (fig. 6, A), is a very distinct and interesting species described by L. S. Strom (1938) from the roots of wild sarsaparilla, *Aralia nudicaulis*, collected at Milwaukee, Wisconsin, on July 2, 1935. A distinctive character is that in 92 out of 104 forewings examined the radial sector is absent. The short cauda and overlapping curved caudal spines are also pronounced.

*The Watercress Aphid.*—The watercress aphid, *Cerosipha roripae* Palmer, is a green and brown species easily characterized by the numerous quite large circular sensoria on antennal segment III of the alates and also by the presence of up to 9 similar sensoria on antennal segment III of the apterae. It has been taken only on watercress, *Roripa hispida*, by M. A. Palmer in Colorado.

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NATURAL SOURCES, HABITATS, AND RESERVOIRS  
OF INSECTS ASSOCIATED WITH  
STORED FOOD PRODUCTS

E. GORTON LINSLEY



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# NATURAL SOURCES, HABITATS, AND RESERVOIRS OF INSECTS ASSOCIATED WITH STORED FOOD PRODUCTS<sup>1</sup>

E. GORTON LINSLEY<sup>2, 3</sup>

INSECTS have long infested man's household and his stored food products, as is evidenced by their presence in Egyptian tombs (Alfieri, 1931)<sup>4</sup> and by references in the Bible (Marlatt, 1896). Yet their place and mode of origin are not well known—an unfortunate fact, since the natural habitats may provide a clue to sources of infestations and other matters of economic import. Apparently no serious attempts have been made to account on this basis for more than a few of the numerous species that infest stored products. (See Marlatt, 1896; Good, 1933; Linsley, 1942a; Hatch, 1942; Hinton, 1943b.) Hatch, in a discussion of the habitat and distribution of some of the more important stored-grain pests, emphasizes the ecological aspects of the problem:

The entomologist . . . sees in these creatures not pests that are competing with humans for the product of their agricultural labors. Rather he finds in them a group of beetles and moths and other insects that at the time when man began the cultivation of wheat had evolved in such a fashion that they fit in with the sort of conditions furnished by the storage and processing of grain. The grain elevator, the flour mill, the grocery store, the housewife's kitchen are simply an enormous insect habitat extending its tentacles into all portions of the wheat-consuming world. The factors governing the occurrence of insects in this vast complex constitute an ecological problem of considerable magnitude.

Entomological literature seldom records the occurrence of stored-products insects in nature. The better the species is known to the economic entomologist, the more is this likely to be true. Since the important stored-products insects are mostly cosmopolitan in distribution, the average collector ignores them and fails to record their capture. Field entomologists have likewise generally ignored them, often assuming that their presence was accidental or resulted from contamination. The present paper will bring together the records<sup>5</sup> available to the writer, together with personal observations, in the hope of clarifying certain problems and of stimulating further research.

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<sup>3</sup> The writer wishes to express his appreciation to Professor O. W. Richards of the Imperial College of Science and Technology, London, and to Dr. H. E. Hinton of the British Museum of Natural History, London, as well as to his colleagues in the Division of Entomology and Parasitology, University of California, for critically reading the manuscript, offering helpful suggestions, and supplying pertinent references.

<sup>4</sup> See "Literature Cited" for complete data on citations mentioned in the text by author and date of publication.

<sup>5</sup> Admittedly some (perhaps many) records derived from the literature may be based on misidentifications. Although this fact may sometimes affect interpretation, it will probably not materially change the conclusions: the latter are based largely on ecological considerations that apply at the generic or group level rather than at the species level. Error might also arise from inaccuracies in the current phylogenetic interpretation of the groups involved. In a few cases, conclusions based on the relatives or congeners of a particular species would perhaps be invalidated by further taxonomic study; some of the supposedly phylogenetic groupings that we now accept may yet prove to have been based on superficial characters.

Stored-products insects undoubtedly share a number of biological characteristics which have combined to bring about their wide distribution and to make them pests of mankind.

One of the most important of these characteristics appears to be a relatively wide range of tolerance for physical factors of the environment, especially moisture. Since in any given environment the extremes may be more significant than the means, these insects can probably tolerate more extreme conditions of temperature and humidity (especially moisture content of food materials) than many less widely distributed forms. Most of the important species can survive, for example, in foodstuffs with a moisture content far below the optimum for their development. Further, although these insects apparently have a broad tolerance for extremes of humidity (and frequently temperature), man maintains optimum conditions for most of the species in buildings, warehouses, and food-storage units.

Second, most stored-products insects have a wide range of food habits, and many include in their diet the majority of dry food and drug products of plant and animal origin. This food range is greater than that generally attributed to most other insects.

Third, nearly all the species involved reproduce more or less continuously under favorable environmental conditions and, with only a few notable exceptions, require no winter diapause. In addition, many of the species are long-lived in the adult stage and may live and reproduce over a period of two or three years.

Fourth, some of the most important species can survive in the adult stage for relatively long periods without food. This fact favors their accidental transportation from one part of the world to another even in the absence of suitable food materials.

Finally, the small size of most of the species may cause them to be overlooked. Frequently their presence remains undetected until a large population has been built up.

## HABITS OF SPECIES IN RELATION TO POSSIBLE SOURCES AND RESERVOIRS

### ACARINA

*Acaridae*.—With the exception of the predaceous and parasitic families (Cheyletidae, Parasitidae, Tarsonemidae, Pediculoididae), most mites associated with stored products are phytophagous. Probably some of these are to be classed as plant scavengers, since they are related to forms which live on corms, roots, and bulbs of living plants. Most of the species, however, are more abundant on stored products when these materials are damp and moldy. This suggests that they feed wholly or partly on molds and other fungi. Several species of *Tyrophagus*, including the sugar mite, *T. longior* (Gerv.), the cereal mite, *T. americanum* (Banks), and a related but unidentified British species (Ewing and Nesbitt, 1942) are recorded as pests of commercial mushroom plantings, a fact which bears out this conclusion. Where direct injury to stored food products has been observed, it has involved mainly the germ area of wheat and other grain.



### THYSANURA

*Lepismatidae*.—The common silverfish, *Lepisma saccharina* Linn., the fire-brat, *Thermobia domestica* Pack., and a few other thysanuran species are known to infest stored products. As food they much prefer starchy materials and cellulose, and their infestation of stored products is largely casual. Both species seek warm locations and are thus rarely found outdoors in the temperate regions, though they have been reported occasionally from under bark, in bird, mammal, and insect nests, in debris, and so forth.

### COLLEMBOLA

*Entomobryidae*.—Two springtails, *Sira buski* Lubbock and *S. platani* Nicolson, have been reported in meal in houses (Mills, 1930), as well as outdoors under bark and in garbage. Apparently they subsist largely on molds and fungi.

### BLATTARIA

*Blattidae*.—Several common roaches, including the oriental cockroach, *Blatta orientalis* Linn., the Australian cockroach, *Periplaneta australasiae* (Fab.), and the American cockroach, *Periplaneta americana* (Linn.), are contaminants or casual pests of stored products. Most of them are omnivorous, feeding on materials of both plant and animal origin. Although they prefer warm habitats, especially heated buildings, they are occasionally found outdoors, even in temperate regions, occurring under bark, in refuse dumps, and the like. Most of the native roaches of the temperate regions are bark inhabitants, but none of these appear to have been recorded from stored food materials.

*Phyllodromiidae*.—Some phyllodromiids, including the German cockroach, *Blattella germanica* (Linn.), and the Surinam roach, *Leucophaea surinamensis* (Linn.), are also casual pests of stored products. In their habits they resemble the true blattids.

### CORRODENTIA

*Cocciliidae*.—*Lachesilla pedicularia* (Linn.) occasionally infests stored food products in Europe and North America. In nature it is found about old wood, straw, ground litter, and plant materials, both living and dead, as well as in birds' nests. It has generally been regarded as a scavenger on plant materials, but may prove to be a fungus feeder.

*Liposcelidae*.—The best-known member of the Liposcelidae is the book louse, or cereal psocid, *Liposcelis divinatorius* (Müll.). It is credited with omnivorous food habits because of the wide variety of plant and animal materials which it infests. Hinton (*in litt.*), however, suggests that this species, as well as most psocids, is probably a fungus feeder. In nature the various species occur under bark, in the nests of birds, mammals, and insects, on fungi, lichens, and algae, as well as in ground debris, straw, and decomposing plant materials—facts which bear out this suggestion.

*Atropidae*.—*Atropos pulsatorium* (Linn.), a common European and North American species infesting furniture, carpets, stored grain, and farinaceous foods, has been found in nature in nests of *Polistes* wasps as well as deserted beehives. Its true food habits are not well known.

## LEPIDOPTERA

*Pyralididae*.—*Pyralis farinalis* (Linn.), the meal snout moth, is a typical feeder on dead and decomposing vegetable materials. It is more common and widely distributed in nature, where it breeds in damp, moldy straw and other plant materials, than it is in stored products. The fact that this species much prefers moldy plant materials and has even been reared from mushroom beds in Virginia (Davis, 1935) suggests that in part, at least, it is a fungus feeder.

*Galleriidae*.—The rice moth, *Corcyra cephalonica* Staint., and a number of its relatives that are found in stored food products are general plant scavengers. It has been found under bark in Australia. But the wax moth, *Galleria mellonella* (Linn.), and the lesser wax moth, *Achroia grisella* (Fab.), are well-known occupants of the hive, where they feed as scavengers on brood combs, frass, pollen, and the like. The lesser wax moth is also said to eat dried apples, raisins, crude sugar, many kinds of seeds, and dried insects. The bumblebee moth, *Aphomia sociella* (Linn.), a common pest of bumblebees' nests in Europe, has occasionally been reported from combs of the honeybee. A related species, *Aphomia gularis* Zell., feeds on many stored products.

*Phycitidae*.—Most of the phycitids are plant scavengers. In this category belong the Indian-meal moth, *Plodia interpunctella* (Hbn.), the Mediterranean flour moth, *Ephestia kühniella* Zell., and other species of *Ephestia*, all of which breed in nature in a wide variety of plant materials (Richards and Thomson, 1932). The tobacco moth, *E. elutella* (Hbn.), the raisin moth, *E. figulilella* Gregs., and the almond moth, *E. cautella* (Walk.), infest dried fruits on the ground in the field as well as in storage.

Since the pests in this family are mostly scavengers, it is not surprising that they can be found breeding in nature under bark or in old wood. *Ephestia elutella* has been found in rotten wood in New York state; *E. cautella* and *E. kühniella* in wood or bark in Europe and North America.

In addition, several species are associated in nature with bees and wasps. *Plodia interpunctella* is a common occupant of bees' nests. Long known as a pest of hives and stored frames (Dunham, 1929), it has also been reported from nests of *Bombus*, *Anthophora*, and *Osmia*. *Ephestia kühniella* is not uncommon in the combs of honeybees and bumblebees as well as social wasps (*Vespa* and *Vespula*). *E. cautella* and *E. elutella* have also been reported from combs of the honeybee. Moths of the genus *Vitula* appear to be definitely associated with social bees (bumblebees and honeybees) and carpenter bees. One of these, the dried-fruit moth, *Vitula serratilineella* Rag., occurs in weak colonies of honeybees and has recently been found breeding in nests of carpenter bees (Linsley, 1943). The closely related *V. edmandsii* (Pack.) often inhabits nests of bumblebees in the eastern United States and has been reported from combs of honeybees in Colorado and Nebraska. It is not yet known to be a pest of stored products.

*Gelechiidae*.—The angoumois grain moth, *Sitotroga cerealella* (Oliv.), is a primary grain pest and infests grain in the field as well as in storage. Since many other species in the same family infest seeds, its food habits are not aberrant. Two related species, *Aristotelia austeropa* Meyr. and *Epitactis studiosa* Meyr., infest stored rice in India (Fletcher, 1916).



*Oecophoridae*.—At least three of the oecophorids occur in stored food products: the seed moth, or brown house moth, *Hofmannophila pseudospretella* (Staint.), and the white-shouldered house moth, *Endrosis lactella* (D. and S.) *Anthonoma xeraula* Meyr. The first two are more or less cosmopolitan; the last is known from India and Japan. They infest dried plant materials, seeds, carpets, clothing, hides, bookbindings, and similar products. In nature they are found in fungi, animal detritus, ground litter, and the like (Morley, 1935, and others). Hinton (1943a) has called attention to larvae of *H. pseudospretella* feeding on dead honeybees near a bees' nest in an old house. He also notes the breeding of this species and one or two of *Endrosis* on dead insects in spider webs under conditions similar to those noted below for certain Dermestidae. The writer has found both *H. pseudospretella* and *E. lactella* breeding in black-widow-spider webs and in house-finch nests.

*Tineidae*.—The members of this family which infest stored products are mostly clothes moths. Concerning the origin of clothes moths, Marlatt (1896) has commented as follows:

The common species of clothes moths have been associated with man from the earliest times and are thoroughly cosmopolitan. They are all probably of Old World origin, none of them being indigenous to the United States. That they were well known to the ancients is shown by Job's reference to "a garment that is moth eaten," and Pliny has given such an accurate description of one of them as to lead to the easy identification of the species. That they were early introduced into the United States is shown by Pehr Kalm, the Swedish scientist . . . who seemed to take a keen interest in house pests. He reported Tineids to be abundant in 1748 in Philadelphia, then a straggling village, and says that clothes, worsted gloves, and other woolen stuffs hung up all summer were often eaten through and through by the worms, and furs were so ruined that the hair would come in handfuls.

What led to the first association of these and other household pests with man is an interesting problem. In the case of clothes moths, the larvae of all of which can, in case of necessity, still subsist on almost any dry animal matter, their early association with man was probably in the role of scavengers, and in prehistoric times they probably fed on waste animal material about human habitations and on fur garments. The fondness they exhibit nowadays for tailor-made suits and expensive products of the loom is simply an illustration of their ability to keep pace with man in his development in the matter of clothing from the skin garments of savagery to the artistic products of the modern tailor or dressmaker.

In the field the clothes moths breed in animal hair, feathers, insect remains, and other dry animal products. Thus Baer (1924) has reported the carpet moth, *Trichophaga tapetzella* (Linn.), from casts (mainly mouse hair, ejected by birds of prey, especially owls); and Laing (1932) from rabbit skins outdoors. One species, the webbing clothes moth, *Tineola bisselliella* (Hüm.), frequents stored food products of animal origin and occasionally plant products that are infested with other insects. The writer has found it breeding in nests of the English sparrow as well as in cells of the bee *Anthophora* in California. Since this species feeds almost exclusively on animal products it probably lives in bees' nests as a scavenger both on insect remains and on stored pollen (which is high in nitrogenous materials).

Several species of *Tinea* also breed under bark, especially when fungus is present (Escherich, 1931). The European grain moth, *Tinea granella* (Linn.), and species of *Setomorpha* have been recorded from this habitat. The writer has found *Tineola bisselliella* breeding under loose bark in southern California. The larvae were apparently feeding on insect remains and detritus.

The so-called cork moth, *Tinea cloacella* Haw., breeds in grain and certain other dried plant products, especially in damp cellars. Since it seeks food of a high moisture content, and occurs in mushrooms (Krause, 1916) and in the fungus *Polyporus* (Morley, 1935), it may be primarily a fungus feeder. Its association with corks of wine flasks is probably due to the presence of molds.

#### COLEOPTERA

*Histeridae*.—At least six species of histerids occur in stored food products although but three of these, *Carcinops 14-striata* Steph., *Dendrophilus punctatus* Hbst., and *Saprinus semistriatus* Scriba, are widespread and common. *C. 14-striata* is a cosmopolitan species found about stored grain and flour, where it is predaceous on grain insects. This species, as well as others in the genus, occurs frequently in rotten wood and under bark. *D. punctatus* is also found in grain, flour, and various stored food products (Cotton and Good, 1937; Ross, 1940). It occurs occasionally under bark in Europe, North America, and probably elsewhere, but most often in nests of ants and of birds (owls, various woodpeckers, crows, the hoopoe, the jackdaw, the starling, screech owls, hawks); also in pigeon and other birdhouses. A related species, *D. opacus* Ross, occurs in nests of woodrats, and *D. tularensis* Ross in those of the ground squirrel. *C. 14-striata* has also been recorded from nests of owls and storks and from birdhouses occupied by starlings and purple martins. *S. semistriatus* Scriba occurs in burrows of the prairie dog.

*Corynetidae*.—The species of *Necrobia* are mainly found associated with decomposing animal matter and oleaginous vegetable materials. Thus the red-legged ham beetle (copra beetle), *Necrobia rufipes* (DeG.), and the red-shouldered ham beetle, *N. ruficollis* (Fab.), often occur in association with *Dermestes*. In the bone rooms of the Museum of Vertebrate Zoölogy of the University of California, where *Dermestes* are used for cleaning skulls, it has not proved practicable to eliminate *Necrobia* completely from the cultures. Both *N. rufipes* and *N. ruficollis*, however, as well as the cosmopolitan *N. violacea* (Linn.), are predaceous or semipredaceous, feeding on other insects (skin beetles, cheese skipper, mites) in the same habitat. This is also true of *Corynetes coeruleus* Dej., a species found about bones, meat, flour, grain, and other products infested with larvae of Dermestidae, Anobiidae, and Ptinidae (Zacher, 1927).

*Dermestidae*.—Characteristic of this group are the skin beetles, *Dermestes* spp., which attack hams and other cured meats, dried fish products, and similar foods of animal origin, and are serious pests in silkworm culture because they injure silk of cocoons. Under natural conditions they breed in dry carcasses of dead animals or in the nests of birds and mammals. Thus *Dermestes bicolor* Fab. occurs in pigeon lofts; *D. elongatus* Lec. in nests of the black-crowned night heron; *D. talpinus* Mann. and *D. signatus* Lec. in those of the magpie, the latter also in crows' nests. In fact, dermestids are among the commonest inhabitants of nests of birds and mammals. In California the writer has frequently encountered the black carpet beetle, *Attagenus piceus* (Oliv.), and the related *Attagenus schäfferi* Hbst. in chipmunk nests; infestations from this source have annoyed many persons in mountain summer cabins. Back and Cotton (1938) have recorded *Attagenus piceus* from birds'



nest; and McAtee (1929) from birdhouses occupied by starlings, English sparrows, purple martins, house wrens, and bluebirds. The related *Attagenus pelli* (Linn.) has also been found in birds' nests. Davis (1934) records *Attagenus elongatulus* Csy. as a regular inhabitant of woodrats' nests in California. *Trogoderma ornatum* (Say) and *T. versicolor* Creutz. have been found in numbers in nests of English sparrows and house wrens; and *Anthrenus occidentis* Csy. in magpie nests. Yokoyama (1929) records adults and larvae of *Anthrenus pimpinellae* (Fab.) in sparrows' nests; and in California *Anthrenus lepidus* and *Attagenus nigripes* Csy. are both common in the nests of the English sparrow, house finch, black phoebe, barn swallow, and cliff swallow.

With few exceptions, the dermestid beetles feed only on products of animal origin. Chittenden (1895a, 1897) and various other authors have called attention, however, to the granivorous habits of a few species, primarily *Trogoderma versicolor* Creutz. and *T. granarium* Everts, but also *Attagenus piccus* and *Anthrenus verbasci* (Linn.).

As might be expected, dermestids inhabit nests of both bees and wasps as well as other insects. Unfortunately the species involved have seldom been carefully identified. Hinton (1934b) has stated that "in practice the nests of Aculeate Hymenoptera are rarely of even slight importance as reservoirs of injurious Dermestidae, and in this respect cannot be compared with either bird (chiefly sparrow) or spider nests which, because of their greater abundance and proximity to houses and warehouses where stored products are kept, are a factor to be reckoned with in the control of these beetles." Possibly Hinton's statement is true for England, but it certainly does not apply to California. In this state, by far the most injurious dermestid to fabrics is *Anthrenus verbasci*; to foodstuffs, *Trogoderma sternale*. In a recent survey the former species was encountered in birds' nests only once and in this case the nest was surrounded by those of *Sceliphron servillei* Sauss. On the other hand, both *Anthrenus verbasci* and *Trogoderma sternale* are common in the nests of *Sceliphron*, *Polistes*, *Odynerus*, and *Osmia*, which regularly nest about buildings as well as in old nests of *Dolichovespula* and *Apis*. In many parts of California where carpet beetles are a serious problem, nests of *Sceliphron* and *Polistes* are far more numerous about buildings than are birds' nests.

According to Hinton (*in litt.*), *Attagenus pantherinus* Ahrens, *Globicornis corticalis* (Eich.), and *Phradonoma villosulum* (Dufts.) all occur in bee or wasps' nests in Europe. Hicks (1926), Mickel (1928), and numerous other authors mention the abundance of *Anthrenus* spp. in nests of *Anthophora*, *Osmia*, and certain other bees. The varied carpet beetle, *Anthrenus verbasci*, is common in wasps' and bees' nests in California; *Anthrenus muscorum* (Linn.) and *Anthrenus (Helocerus) fuscus* (Latr.) breed in the nests of *Osmia* or other bees in Europe; and Hinton (1943a) has recorded *Attagenus pelli* from the vicinity of honeybees' nests in England. In California *Attagenus ?piccus*, *Trogoderma ajar* (Csy.), and *T. sternale* Jayne occur in the nests of *Anthophora* and *Osmia*. *Trogoderma glabrum* Hbst. breeds in the nests of *Osmia* and *Odynerus* in Europe; the larger cabinet beetle, *T. versicolor*, in bees' and wasps' nests in North America; and the common *T. ornatum* has been recorded from honeycomb and from the nests of spiders. Another species, *T. socium* Lea, was originally described from ants' nests in Sydney. *Mcgatoma*

*undata* (Linn.) also lives with bees in Europe, and *Perimegatoma vespulae* Milliron was bred originally from the nests of *Dolichovespula arenaria* (Fab.) in Minnesota. Riley (1870), Lintner (1890), and others have noted the feeding of the larder beetle, *Dermestes lardarius* Linn., in combs of the honeybee; and Milum (1940b) discusses Dermestidae feeding on brood combs containing pollen, dead larvae, or bees but does not specify the species involved.

It may be significant that at least four species of *Trogoderma* live in nests of bees or wasps. In California, *T. ajax* can be reared entirely upon a diet of pollen, but will also consume animal products (insect parts) and grain. The larvae of both *Attagenus piceus* and *Anthrenus verbasci* also feed on cereal products (Griswold, 1941). Both these species occur in bees' nests and in the larval stage may be reared on pollen. Most of the species of *Attagenus* and *Anthrenus* feed as adults upon pollen and nectar, which they obtain directly from flowers. Possibly, therefore, pollen may have influenced the evolution of food habits of certain small dermestids associated with stored food products.

Dermestids are also commonly associated with nests and webbing of various caterpillars. In Japan and elsewhere they are among the most important pests of the silk industry. The breeding of *Dermestes* in nests of processionary and tent caterpillars has been noted by several writers (Perris, 1853; Bristowe, 1928, and others). Hinton (1943b) summarizes these as follows:

In Europe a few Dermestidae are known to breed in the nests of certain gregarious caterpillars of the families Thaumetopoeidae and Lymantriidae. *Dermestes mustelinus* Er., *D. aurichalceus* Kuster, *D. lardarius* L., and *Anthrenus vorax* Waterh. are found in the nests of *Thaumetopoea pityocampa* Schiff. *D. aurichalceus* has also been recorded in the nests of *T. processionea* L. *D. erichsoni* Ganglb. and *D. tessellatus* F. have been found in the nests of the brown-tail moth, *Euproctis chrysorrhoea* L. and the former also in the nests of *Euproctis similis* Fuess. The larvae and adults of the species of *Dermestes* feed on the dead or injured moth larvae. *Dermestes mustelinus* is said to be of some value in controlling *T. pityocampa* in Catalonia.

Strickland (1929) has recorded larder-beetle infestations in Canada arising from nests of tent caterpillars. Donohoe (1939) has also called attention to the abundance of dermestids in webbing of *Ephestia* in raisin storage. He found thousands of larvae of *Trogoderma versicolor* Cz. breeding in webbing under stacks of the dried fruit. *Trogoderma sternale* Jayne and *Perimegatoma variegatum* Horn were also found in the same habitat.

A related habitat which may provide sources of household and warehouse infestation in spiders' sheet-webbing, which provides sources of dead insects (Auten, 1925; Hinton, 1943b, and others). In California both *Anthrenus verbasci* and *Trogoderma sternale* breed in this habitat. Hinton has summarized the published records as follows:

Seven species of Dermestidae have been recorded in or near spider webs, *Attagenus pellio* L., *Ctesias serra* Steph., *Megatoma undata* L., *Globicornis marginata* Payk., *Trogoderma ornata* Say, *Anthrenus museorum* L., and *Trinodes hirtus* F. A further species *Anthrenus fuscus* Oliv., has recently (vi-xi. 1942) been found by me to be very common in Linton and Little Abington, Cambs. about stone walls, sheds, barns, and houses in or about the webs of *Tegenaria domestica* L. and other spiders. *A. museorum* frequently occurred in the same webs. The larvae of both species of *Anthrenus* are also very common in spider webs about sheds and outhouses in Reading (*A. S. Corbet*) and Slough (*O. W. Richards*). A few



larvae of both species were found by me during the summer of 1942 under bark in Linton with the spiders *Clubiona corticalis* Walek., *Ciniflo ferox* Walek., *Harpactea hombergii* Scopoli and *Segestria semoculata* L. The breeding habits out of doors in Britain of these two species of *Anthrenus* [do] not appear to have been previously known.

Many of the household and food-infesting dermestids are also common under bark, where they feed on debris of animal origin. This is especially true of the smaller species (*Trogoderma*, *Anthrenus*, *Attagenus*). Many authors have recorded instances of *Dermestes* attacking wood; but Walker (1884), Richards (1931), and others have pointed out that attacks of dermestids on certain plant materials (wood, cork, tobacco, bales of cotton, and books, as well as such inorganic materials as salt, plaster, asbestos, and lead) are due to larvae seeking a place for pupation. Patton (1931) illustrates injury to wood by pupating larvae of the hide beetle, *D. vulpinus* Fab.

Among the dermestids common under bark and in old burrows of wood borers are species of *Trogoderma*. *T. ornatum* breeds in this habitat in many localities in eastern North America and perhaps also in California. (Records from this state are possibly the result of misidentification.) Auten (1925) records it as feeding on spiders' eggs under bark in Ohio. *T. sternale* occurs under bark in San Diego County, California; and another species (identity uncertain) is not uncommon in the same habitat in the Sierra Nevada and northern California. The same species or a closely related one has also been found in Colorado under bark of ponderosa and limber pine (De Leon, *in litt.*). *T. versicolor* has been reared from dead hickory in New York, as well as from woolens and stored grain. Exact records of the occurrence of *Anthrenus* and *Attagenus* are rare because of uncertainties of identification. Both genera breed, however, in the bark habitat; and at least two species of the former and one of the latter occur under bark of various trees in California. The skin beetles, *Dermestes* spp., have also been found under bark from time to time, but probably mainly during hibernation (Blatchley, 1910).

*Ostomatidae*.—Nearly all ostomatids are predators living under bark or in the galleries of wood-boring insects. The species of *Temnochila* are characteristic of the group, spending both the adult and the larval stages as predators on wood-boring insects. One species, however, *T. coerulea* (Oliv.), occurs also in granaries (Zacher, 1927; Cotton and Good, 1937), where it is presumably predaceous on grain-feeding insects.

The species of *Tenebroides* are also typical inhabitants of bark and wood. Both as larvae and as adults, they live under bark as semipredators. When ready to hibernate or pupate, the larvae bore into the bark and construct a resting cell. At least three species of *Tenebroides*, however, are found in stored food products. The best-known is the cadelle, *T. mauritanicus* (Linn.). This pest probably does as much harm by boring into wood and other food containers for hibernation and pupation as by injuring or contaminating food. The adults are predaceous on their own larvae as well as on those of *Plodia interpunctella*, *Oryzaephilus surinamensis* (Linn.), and other food-inhabiting insects. The larvae feed on broken grain and certain other plant products, but are also semipredaceous and on some materials cannot complete development without supplementing their diet with dead or living insects. The cadelle has been found living under bark in both Europe and North America. The nor-

mally bark-inhabiting species *T. nanus* (Melsh.) and *T. corticalis* (Melsh.) both also infest stored grain in North America (Cotton and Good, 1937). The latter is also a predator of codling-moth larvae (Woodside, 1942). A related species, *T. maroccanus* Reitt., is predaceous on gypsy-moth eggs (de Lepiney, 1933).

The Siamese grain beetle, *Lophocateres pusillus* (Klug), infests a wide variety of stored foodstuffs in tropical and subtropical regions. It has occasionally been found under bark in nature.

*Nitidulidae*.—The most important members of this family, the dried-fruit beetles, *Carpophilus* spp., have come directly from forms that naturally feed either on dry or rotting fruit or upon oozing sap, pollen, fungi, or other plant materials. *Haptoncus luteolus* (Er.) lives in the field on similar materials and has been reported from mealybug-infested sugar cane (Swezey, 1942). It infests granaries and dried-fruit warehouses in California and elsewhere. *Nitidula ziczac* Say has sometimes infested hides in California. In nature it is found on dead birds and reptiles (Blatchley, 1910). A few species also occur in nests of bees. In Australia, *Carpophilus planatus* Mur., *Brachypeplus auritus* Mur., and *B. basalis* Er. occur in nests of *Trigona*; *B. inquilinus* Lea and *B. blandus* Mur. in wild nests of the honeybee.

*Silvanidae*.—The silvanids are common in nature beneath bark. Various species of *Silvanus* have been noted in this habitat by many authors. The saw-toothed grain beetle, *Oryzaephilus surinamensis*, and the merchant grain beetle, *O. mercator* (Fauv.), both may occasionally breed under bark, especially in tropical and subtropical regions. The same is true of the square-necked grain beetle, *Cathartus quadricollis* (Guer.), and the foreign grain beetle, *Ahasverus advena* (Watl.). Lucas (1849) found the latter species under cork-oak bark in North Africa; and Fall (1901) found it, possibly hibernating, under eucalyptus bark in southern California in December. *C. quadricollis* feeds on grain and farinaceous materials, and infests wheat and corn in the field as well as in storage. *Ahasverus advena* is commonly found on grain in the field and on stored grain and milled products which are slightly out of condition (Cotton and Winburn, 1941). It probably feeds on molds and fungi. The food habits of *Monanus concinnulus* Walk. and *Nausibius clavicornis* Kug. are not well understood. The former has been recorded in bran and flour in Mauritius; the latter is common among ships' stores and tropical cargoes, especially sugar. It may feed largely on molds and fungi.

*Cucujidae*.—Most of the cucujids live in nature under bark of trees. This is true of the flat grain beetles (*Laemophloeus* spp.); and, in fact, the generic name *Laemophloeus* means "bark eater." Wheeler (1921) summarizes the habits of twenty species, most of which live under bark in association with bark beetles (Scolytidae). The rust-red grain beetle, *L. ferrugineus* (Steph.), has often been recorded from under bark as well as in the nests of *Vespa*. Shepard (1936) observed that it is at least to some extent predaceous on other insects. He found the oviposition rate highest when females were fed only on eggs of the Angoumois grain moth, and larval development most rapid when insects were available as food. Several other bark- and wood-inhabiting species are recorded as contaminants of stored food products: *L. ater* (Oliv.) and *L. emgei* Reitt. in Europe; *L. janeti* Grouv. in England, Africa, and the Orient;



and *L. turcicus* Grouv. in many parts of the world. Glover (1869) has recorded *L. modestus* Say from stored hempseed in the United States.

One additional bark-inhabiting cucujid is a stored-products pest. *Laemotmetus rhizophagoides* (Walk.) is found in stored rice in Africa, the Indo-Malay and other Asiatic regions, and certain parts of Europe.

*Cryptophagidae*.—Approximately nineteen species of *Cryptophagus* and a half-dozen other members of this family—such as *Henoticus californicus* Mann. and *Pharaxonotha kirschi* Reitt.—have been recorded from houses, warehouses, mills, and granaries. *Cryptophagus distinguendus* Sturm has been recorded from an edible fungus (*Psalliota*) by Ripper (1930); and Hinton and Stephens (1941) have reared *C. acutangulus* Gyll. on a wide variety of fungi, finding that the species would thrive on a diet of *Penicillium* conidia. *C. acutangulus* also proved to be capable of mechanical, internal, and external transportation of fungi. Other cryptophagids probably have habits similar to these and feed almost entirely on fungi and molds rather than the stored food products in which they are found. In nature they occur under bark; in nests of birds, mammals, and insects; and in fungi, ground debris, straw, and decomposing plant materials.

At least three genera of cryptophagids have been found in nests of bees or wasps. Many of the species of *Cryptophagus* recorded from buildings have sometimes been found living with social bees and wasps in Britain. *C. badius* Sturm occurs with the giant hornet, *Vespa crabro* Linn.; *C. pubescens* Sturm, *C. pilosus* Gyll., and *C. distinguendus* with one or more species of *Vespula*; and *C. punctipennis* Bris., *C. umbratus* Er., and *C. saginatus* Sturm with *Bombus*. *C. distinguendus* has also been found in bumblebee nests, as has *C. croceus* Zimm. Faleoz (1929–1931) has recorded the well-known *C. cellaris* (Scop.) from beehives in Europe.

*Atomaria ruficornis* Marsh. and *Atomaria pellata* Kraatz have occurred in nests of *Vespa*. *Antherophagus pallens* (Fab.) and *Antherophagus nigricornis* (Fab.) are common in bumblebees' nests in Europe; and *Antherophagus ochraceus* Melsh. in similar situations in the eastern United States. Cryptophagids have also been found in nests of processionary caterpillars (Bristowe, 1928, and others).

*Mycetophagidae*.—Several mycetophagids may also be found in stored products. The hairy fungus beetle, *Typhaca stercorica* (Linn.), is the best known of these. *Mycetophagus quadriguttatus* Müll., *M. bipustulatus* Melsh., and *Litargus balteatus* Lee. are less common. All have been recorded from fungi in nature, and their food habits probably resemble those of the preceding family.

*Lathridiidae*.—Thirty or more species of lathridiids have been recorded from stored food products, warehouses, granaries, or mills. *Corticaria serrata* Payk. has also been found in mushroom beds (Davis, 1935); and Hinton (1941b) has shown that both larvae and adults of the species studied by him feed on mycetozoa and fungi, particularly molds.

Thus it is not surprising that various species occur under bark, where many kinds of fungi are available. Of the lathridiids recorded from stored food products, *Holoparamceus caularum* (Aubé), *H. depressus* Curtis, *H. singularis* (Beck), *Coninomus constrictus* (Gyll.), *Coninomus nodifer* (Westw.),

*Enicmus minutus* (Linn.), and *Corticaria fenestralis* (Linn.) have all been found commonly under bark.

Many lathridiids also regularly live with bees, wasps, and ants. Among the stored-products species recorded from this habitat are *Coninomus constrictus* from ants' nests (*Lasius* spp.); and *C. nodifer* and *Enicmus minutus* from nests of bumblebees (*Bombus* spp.) and wasps. (*Vespa* spp.).

Lathridiids also often inhabit the nests of birds and mammals. *Enicmus crenatus* Lec., *E. suspectus* Fall, *Metophthalmus trux* Fall, all occur in nests of wood rats (*Neotoma*) at Berkeley, California. *E. minutus* and *Cartodere ruficollis* (Marsh.) have been recorded from storks' nests in Europe, and *Corticaria pubescens* (Gyll.) from birds' nests in England. But since lathridiids are primarily fungus feeders, their occurrence in animal nests indicates little more than the presence of suitable molds.

*Mycetaeidae*.—The hairy cellar beetle, *Mycetaea hirta* Marsh., is found in Europe and North America. It has been recorded from fungi, stored grain, wine cellars, and corks. According to Hinton (*in litt.*) fungus is its primary, if not exclusive, food. Its presence on corks is undoubtedly due to fungi or molds.

*Tenebrionidae*.—The stored-products pests of the family Tenebrionidae belong, with few exceptions, to the Ulominae. Most members of this subfamily live under bark or in rotting wood as semipredators and scavengers. In the United States, native species of *Uloma* are common in this habitat. In Europe, *Uloma culinaris* (Linn.) is said to be injurious in granaries and other places of food storage (Zacher, 1927; Cotton and Good, 1937). In nature it lives under bark and in burrows of bark beetles and wood borers.

The broad-horned flour beetle, *Gnathocerus cornutus* (Fab.), occurs under bark in California (Chittenden, 1895*b*; Essig, 1926) and North Africa (Lucas, 1849). The slender-horned flour beetle, *Gnathocerus maxillosus* (Fab.), is not uncommon under bark in tropical and subtropical regions. Wollaston (1860) noted it from under bark of plane trees in Madeira, and it has also been recorded from under bark of poplar in Europe. De Leon (*in litt.*) found it abundant under bark of *Pistacio simarubra* in Puerto Rico; and the writer took it under bark in Java.

*Cynaesus angustus* (Lec.) has been recorded from flour mills in Washington state (Hatch, 1940), and shelled corn in Minnesota. Both this species and the related *C. depressus* Horn are native to western North America. In nature they breed under bark, or about the base of yucca plants in Arizona and southern California.

The long-headed flour beetle, *Latheticus oryzae* Waterh., also lives in rotten wood and under bark. Blair (1930) records it in India as having been bred from wood, where it may have fed on remains of wood borers. A closely related native American species, *L. prosopis* Chitt., breeds regularly under bark of dead mesquite in the southwestern United States.

The various species of *Tribolium* are also to be found under bark (Good, 1936). The rust-red flour beetle, *T. castaneum* (Hbst.), is occasionally found in this habitat in Britain and the United States; more frequently in the Dutch East Indies, the Malay region, and India. According to Stebbing (1914) it preys upon the small bamboo shot borer, *Dinoderus minutus* (Fab.). The confused flour beetle, *T. confusum* duV., occurs under bark in California and

parts of southern and southwestern United States. Stebbing recorded it from India as a semipredator on the bamboo shot borer, *Dinoderus pilifrons* Lesne. Many have noted its semipredaceous behavior in stored food products. In Europe and North America the black flour beetle, *T. madens* Charp., lives regularly under bark. De Leon (*in litt.*) records it from ponderosa and limber pine in Colorado, and it has been found breeding in similar habitats elsewhere. Leech (1943) found it in beehives in British Columbia as well as under boards and stones. Adults were observed to kill larval Bibionidae in leaf mold under a log. *T. indicum* Blair, which lives under bark in India (Blair, 1930), has not yet been recorded from stored food products.

The small-eyed flour beetle, *Palorus ratzeburgi* Wissm., and the depressed flour beetle, *P. subdepressus* Woll., have both been found under bark in Europe; the former also in Guam. *P. depressus* (Fab.) apparently breeds regularly in dead wood and under bark. In grain and flour it appears to be less common than the two other species. Blair (1930) has recorded nine additional species of *Palorus* from on or under the bark of trees infested with borers in India, presumably the original home of the group.

The false flour beetles (*Aphanotus* spp.) are common bark inhabitants in western North America. *A. parallelus* Csy. breeds under bark in Arizona and southern California and has also been found in various stored food products. The increasingly injurious *A. destructor* (Uytt.) occurs under bark in central California, and *A. brevicornis* (Lee.) in many parts of the Pacific Coast. The latter, however, is primarily associated with wood-nesting bees (*Xylocopa*).

The lesser mealworm, *Alphitobius diaperinus* (Panz.), was once found by the writer in large numbers under bark of a dead oak tree near Crockett, California. It occurs most commonly in moldy flour. According to Tischler (1937), the adults also feed on dead insects; but they are perhaps not predaceous. The grain mold beetle, *A. piccus* (Oliv.), has also been found under bark in southern California and in India. Both species probably subsist largely on molds and fungi.

*Sitophagus hololeptoides* (Cast.) breeds under bark in Central America. It has also been recorded from cereals and other stored products (Chittenden, 1911a; Cotton and Good, 1937).

The species of *Doliema* and *Platydemia* also regularly live under bark. Thus Blatchley (1910) records *D. pallida* (Say) from this habitat in Indiana, and Fall (1901) states that *D. plana* (Fab.) is common in southern California under bark of sycamores. P. C. Ting reports that the latter is commonly intercepted in quarantine from under bark of lignum vitae logs from Mexico, Nicaragua, Guatemala, and Costa Rica. *P. ruficornis* Sturm, a common native bark inhabitant, has attacked shelled corn and shorts in Kansas (Cotton and Good, 1937).

The species of *Tenebrio* may also be found in old wood and under bark. The commonest species in this habitat is *T. picipes* Hbst., which occurs under bark of various trees from coast to coast. The related *Idiobates castaneus* (Knoch) occupies the same habitat.

Many bark-inhabiting tenebrionids are fungus feeders. Most of the species found in stored food products will probably feed on any fungi available. Several, including the two-banded fungus beetle, *Alphitophagus bifasciatus*



(Say). *Alphitobius piceus* (Oliv.), and *Alphitobius diaperinus*, appear to be primarily fungus feeders and are found in food products only when these materials are in a somewhat deteriorated condition.

Several tenebrionids inhabit bees' nests. The false black flour beetle, *Aphanotus destructor*, occurs in the nests of *Osmia* and *Anthophora*; and the closely related *Aphanotus brevicornis* breeds regularly in cells of *Xylocopa* and has also been recorded from those of *Anthidium*. Another flour beetle, *Cynaesus angustus*, has been found in *Osmia* nests in southern California. *Tribolium castaneum* has also been reported from nests of *Megachile* in Hawaii; *Tribolium confusum*, from cells of *Anthophora*, *Osmia*, and *Clisodon* in California. Frison (1926) has recorded two species of mealworms, *Tenebrio obscurus* Fab. and *Tenebrio tenebroides* Beauv., from *Bombus* nests in Illinois. In Australia, Lea (1905) has described *Tribolium myrmecophilum* Lea from the nests of ants, and *Palorus eutermiphilus* Lea from those of termites.

*Ptinidae*.—The ecology and habits of ptinids in nature are not well understood. A few species, however, occur under bark or in rotten wood. Among the economic species, Fowler (1890) has noted in old wood the white-marked spider beetle, *Ptinus fur* (Linn.), the brown spider beetle, *P. hirtellus* Sturm, and *P. subpilosus* Sturm. The white-marked spider beetle has also been found under bark in Ohio (Auten, 1925). Hinton (1941a) records *P. rufipes* Oliv. from dead branches and *P. palliatus* Perris from old posts and similar places. According to Picard (1919), the larvae of *P. lichenum* Marsh. live in the dry wood and bark of fig trees. Blair (1930) records *P. pusillus* Sturm from a rotten staircase, and De Leon (*in litt.*) has reported *Ptinus* sp. from under bark of limber pine. A native bark- and wood-inhabiting species, *P. gandolphei* Pic, has been reported from raisins in California (Donohoe, 1939). Finally, Stebbing (1914) has noted *Gibbium* sp. living under the bark of teakwood.

More important, however, is the fact that several ptinids are found in nests of birds and rodents. *Tipnus unicolor* Pill. and Mitt. occurs in nests of the house martin and other birds; *Ptinus sexpunctatus* Panz. in those of the house martin; *P. hirtellus* Sturm in those of the English sparrow; *P. fur* in robins' nests and in birdhouses occupied by house wrens and bluebirds; and *P. agnatus* Fall in nests of a woodrat (*Neotoma*). The writer has found *P. gandolphei* breeding in nests of the barn owl, and on several occasions *Mezium affine* Boield has been found in nests of Norway rats in San Francisco. Hinton (*in litt.*) notes that many species feed on rodent dung, a fact which may account for their occurrence in nests of birds and mammals. Such nests, however, frequently contain other plant and animal products acceptable to ptinids.

Ptinids also occur in nature in nests of other insects. *Ptinus sexpunctatus* was apparently the first ptinid species taken in association with bees. In 1872, Bebel reported its capture at the nest entrances of *Megachile muraria* (Retz.). It has since been recorded from nests of the following bees: *M. muraria*, *M. willughbiella* (Kirby), *Osmia rufa* (Linn.), *O. cornuta* (Latr.), *O. emarginata* Lep., *O. villosa* (Schenck), *O. mitis* Nyl., *Hoplitis adunca* (Panz.), *Chelostoma nigricorne* (Nyl.), and possibly also *Bombus* sp. The well-known white-marked spider beetle, *Ptinus fur*, also feeds on pollen in bees' nests. It has been recorded from *O. tridentata* Duf. and Per., *O. rufa*, and *C. maxillosa* (Linn.), as well as from the combs of the honeybee, *Apis mellifera* Linn., and

the giant hornet, *Vespa crabro*. Cros (1932) describes *P. vaulogeri* Pic living in the nests of *O. longispina* Perez; and Linsley and MacSwain (1942a) have discussed the California spider beetle, *P. californicus* Pic, as a depredator in nests of *O. lignaria* (Say) and *Anthophora linsleyi* Timb. Arnhart (1929) and Brassler (1929) have also called attention to depredations of *P. raptor* (Linn.) on pollen stores of honeybees in Europe; and Lea (1905) records *P. exulans* Er. from deserted hives in Australia. The latter author has also reported a long series of ptnids (*Polyplocotes*, *Diplocotes*, *Diphobia*, *Paussoptinus*, *Hexaplocotes*, *Ectrephes*) that appear to live regularly in nests of ants of the genus *Iridomyrmex*. In Europe the golden spider beetle, *Niptus hololeucus* (Fald.), has been reported from nests of the wasp *Dolichovespula*, and in California *P. hirtellus* breeds in nests of *Sceliphron servillei*.

*Anobiidae*.—The drugstore beetle, *Stegobium paniceum* (Linn.), and the tobacco beetle, *Lasioderma serricorne* (Fab.), are principally scavengers on dead plant products, but were doubtless derived originally from wood-boring ancestors, for other anobiids are notorious wood borers. The literature often reports the tobacco beetle as occurring under bark or boring in woods; but there is some question about the identification of the species. The writer has found the drugstore beetle boring in bamboo mah-jongg playing pieces, where the larvae fed also on the glue supporting the ivory face; and in thin wood forms of food containers. Linsley and MacSwain (1942b) have noted the drugstore beetle feeding on pollen in the nests of *Anthophora linsleyi* in southern California. Rayment (1935) states that the tobacco beetle lives in the cells of *Anthophora* in Australia; but his figures (pl. 45, figs. 19, 21) are of the drugstore beetle, a much more likely nest inhabitant.

*Bostrichidae*.—This family is composed of true wood borers. A few species have, however, transferred from a wood-boring to grain-boring habit. The best known is the lesser grain borer, *Rhyzopertha dominica* (Fab.). It must also be classed as a seed-infesting species that has been carried over from field to stored products. Gurney (1918) actually records it from wheat ears in the field. Nevertheless, the species must have been derived originally from a wood-boring type. Potter (1935) writes as follows:

There seems little doubt that *R. dominica* originally fed solely on wood, probably living wood. Lesne . . . records having seen a pair working at a communal gallery on a tree, probably *Quassia* spp., in French Guinea, and Lucas . . . has recorded it under the bark of *Quercus suber* and *Cytisus spinosus*. More recently Gardner . . . records a long series bred from wood of *Artocarpus hirsutus*. The species of tree or shrub that was the original host of *R. dominica* cannot definitely be ascertained, since, in addition to the above records, it is reported as attacking various unidentified woods.

Experimentally, Potter induced the species to attack young maize plants in confinement; also the soft inner cane and pith of bamboo. It has attacked cork in insect boxes (Riley, 1882, and others), wood of sugar casks (Gorham, 1883), wooden bins and containers for wheat and other grains (Mackie and Carter, 1937), and wooden packing cases (Froggatt, 1918). As a grain feeder it retains the boring habit, the young larva tunneling into the grain, the larva feeding and pupating within the kernel, and the adult boring its way out upon maturity. A related species, *R. hordecum* Mats., has been recorded from stored grain and plant products in Formosa (Clausen, 1931).

The genus *Dinoderus* contains several species that bore in bamboo. Two of these are known to infest grain, spices, and drugs. The small bamboo shot borer, *D. minutus*, a well-known bamboo pest, has been recorded from spices, cacao, and the like by Richards and Herford (1930); and from stored grains, flour, tobacco, and nuts by other authors. *D. bifoveolatus* Woll. has been found in similar products in England (Laing, 1929).

The larger grain borer, *Protostephanus truncatus* (Horn), a species related to *Dinoderus*, also infests stored grains and tubers (Chittenden, 1911b).

*Bruchidae*.—Practically all bruchids are seed-infesting species in nature, and most of them are restricted to seeds of legumes. Of these, a great many infest stored seeds, including the bean weevil, *Acanthoscelides obtectus* (Say), the spotted bean weevil, *Callosobruchus maculatus* (Fab.), the Mexican bean weevil, *Zabrotes pectoralis* Sharp, and several other species which attack beans and peas both in the field and in storage. Certain others, including the pea weevil, *Bruchus pisorum* (Linn.), cannot reproduce in stored beans and peas and are classed as pests of stored food products only because they emerge from material previously infested in the field.

*Curculionidae* and *Platystomidae*.—The weevils, particularly the granary weevil, *Sitophilus granarius* (Linn.), and the rice weevil, *S. oryza* (Linn.), were obviously derived from types infesting grain in the field. Although both appear to be natives of the Old World, their geographical or topographical origins may not have been identical, since the former prefers the more temperate regions north of 40 degrees latitude, the latter the regions southward. Other true weevils, including the broad-nosed grain weevil, *Caulophilus latinasus* (Say), and platystomids such as the coffee-bean weevil, *Araecerus fasciculatus* (DeG.), must have had a similar origin, since nearly all of them infest seeds in the field as well as in storage.

*Scolytidae*.—The scolytids are mainly bark beetles, only a few species being classed as true wood borers. Three species, all belonging to the genus *Pagiocerus*, have been recorded as damaging corn in Central and South America (Cotton and Good, 1937). They are obviously derived from wood-boring types.

## DIPTERA

*Piophilidae*.—Few Diptera other than fruit flies and muscoid scavengers are true pests of stored products. The best-known species, the cheese skipper, *Piophilidae casei* (Linn.), feeds on moist animal products, especially those that are high in fat content (such as cheese, bacon, ham). In nature it breeds in excrement, dead animals, and similar materials.

## DISCUSSION

### NATURAL SOURCES OF STORED-PRODUCTS INSECTS

As we have seen, insects associated with stored food products have come from a variety of habitats. They may be classified according to their sources as follows: (1) seed-infesting species; (2) fungus-feeding-species; (3) scavengers on dead plant materials; (4) scavengers on dead animal materials; (5) scavengers or semipredators living under bark; (6) wood borers and wood scavengers; (7) scavengers or depredators in the nests of other insects and



spiders; (8) parasites and predators. Naturally, there is a considerable overlap; some species belong in two or more categories.

*Seed-infesting Species.*—Of the many insects that infest seeds under natural conditions, a number will attack seeds stored by man. Such behavior involves little change in food habits and probably little adjustment to a new environment, although some change in tolerance of relative humidity and moisture content of food was probably required in many cases. The surprising thing, perhaps, is that so few species of stored-food insects, except the coleopterous family Bruchidae, are of this sort. Among the few involved, however, are some serious pests. Certain of these are still to be found infesting the flower head or the seed pod; others the seeds that have been shed on the ground. A few species avail themselves of the seed storages of birds and mammals and have had to make only a small transition to the storehouses of man. The following list includes the better-known seed-infesting species:

Lepidoptera	Coleoptera, <i>continued</i>
Gelechiidae	Platystomidae
<i>Sitotroga cerealella</i> (Oliv.)	<i>Araecerus fasciculatus</i> (DeG.)
Coleoptera	<i>Brachytarsus alternatus</i> (Say)
Bruchidae	<i>Brachytarsus sticticus</i> (Boh.)
<i>Acanthoscelides obtectus</i> (Say)	Curculionidae
<i>Callosobruchus maculatus</i> (Fab.)	<i>Sitophilus granarius</i> (Linn.)
<i>Zabrotes pectoralis</i> Sharp	<i>Sitophilus oryza</i> (Linn.)
Etc.	<i>Caulophilus latinasus</i> (Say)

*Fungus-feeding Species.*—Numerous species of insects which occur in stored food products are primarily fungus feeders. They are most commonly found in old, damp products and probably confine their feeding to the fungi and molds present. Included are beetles of the families Cryptophagidae, Mycetophagidae, and Lathridiidae, of which many species have been found in grain and grain products. Some of the Tenebrionidae, and some moths and mites also belong wholly or partly in this category.

Acarina	Coleoptera, <i>continued</i>
Acaridae	Lathridiidae
<i>Acarus siro</i> Linn.	<i>Holoparamesus depressus</i> Curtis
<i>Carpoglyphus passulorum</i> Her.	<i>Metophtalmus serripennis</i> (Broun)
<i>Tyrophagus americanum</i> (Banks)	<i>Coninomus nodifer</i> (Westw.)
<i>Tyrophagus putrescentiae</i> (Schr.)	<i>Enicmus minutus</i> (Linn.)
Etc.	<i>Adistemia watsoni</i> (Woll.)
Collembola	<i>Cartodere filum</i> (Aubé)
Entomobryidae	<i>Cartodere filicornis</i> (Gyll.)
<i>Sira buski</i> Lubb.	<i>Cartodere ruficollis</i> (Marsh.)
<i>Sira platani</i> Nie.	<i>Corticaria fulva</i> (Com.)
Lepidoptera	Etc.
Tineidae	Mycetophagidae
<i>Tinea cloacella</i> Haw.	<i>Mycetophagus bipustulatus</i> Melsh
Pyrallidae	<i>Mycetophagus quadriguttatus</i> Müll.
<i>Pyralis farinalis</i> (Linn.)	<i>Typhaea stercora</i> (Linn.)
<i>Pyralis lienigalis</i> Zell.	<i>Litargus balteatus</i> Lec.
<i>Hypsopygia costalis</i> (Fab.)	Mycetaeidae
Coleoptera	<i>Mycetaea hirta</i> Marsh.
Cryptophagidae	Tenebrionidae
<i>Henoticus californicus</i> Mann.	<i>Alphitophagus bifasciatus</i> (Say)
<i>Cryptophagus acutangulus</i> Gyll.	<i>Alphitobius piceus</i> (Oliv.)
<i>Cryptophagus</i> spp.	<i>Alphitobius diaperinus</i> (Panz.)

*Scavengers on Dead Plant Materials.*—Insect scavengers of plant materials provide an obvious source for stored-food-products pests. A few of the many species involved are listed below.

Thysanura	Lepidoptera
Lepismatidae	Galleriidae
<i>Lepisma saccharina</i> Linn.	<i>Corcyra cephalonica</i> Staint.
<i>Thermobia domestica</i> Pack.	Phycitidae
Blattaria	<i>Ephestia cautella</i> (Walk.)
Blattidae	<i>Ephestia elutella</i> (Hbn.)
<i>Blattella germanica</i> (Linn.)	<i>Ephestia figulilella</i> Gregs.
<i>Blatta orientalis</i> Linn.	<i>Ephestia kühniella</i> Zell.
<i>Periplaneta americana</i> (Linn.)	<i>Plodia interpunctella</i> (Hbn.)
<i>Periplaneta australasiae</i> (Fab.)	Coleoptera
<i>Leucophaea surinamensis</i> (Linn.)	Nitidulidae
Corrodentia	<i>Carpophilus dimidiatus</i> (Fab.)
Caeciliidae	<i>Carpophilus hemipterus</i> (Linn.)
<i>Lachesilla pedicularia</i> (Linn.)	<i>Carpophilus decipiens</i> Horn
Liposcelidae	<i>Carpophilus ligneus</i> Mur.
<i>Liposcelis divinatorius</i> (Müll.)	<i>Carpophilus pallipennis</i> Say
Atropidae	<i>Carpophilus</i> spp.
<i>Atropos pulsatorium</i> (Linn.)	<i>Urophorus humeralis</i> (Fab.)

*Scavengers on Dead Animal Materials.*—Here again we find many insects that have carried over their preference for dead animal materials in the field to products of a similar origin stored by man.

Coleoptera	Lepidoptera
Dermestidae	Tineidae
<i>Dermestes</i> spp.	<i>Trichophaga tapetzella</i> (Linn.)
<i>Attagenus</i> spp.	<i>Tineola bisselliella</i> (Hüm.)
<i>Anthrenus</i> spp.	<i>Tinea</i> spp.
<i>Perimegatomia</i> spp.	<i>Setomorpha insectella</i> (Fab.)
<i>Trogoderma</i> spp.	<i>Setomorpha margaloestriata</i> Keuch.
Nitidulidae	Diptera
<i>Nitidula ziczac</i> Say	Piophilidae
	<i>Piophila casei</i> (Linn.)

*Scavengers or Semipredators Living under Bark.*—Many pests of stored food products appear to have been derived from forms that live under bark, the majority coming originally from tropical and subtropical regions. The bark habitat, aside from offering protection and favorable physical conditions, provides varied foods of both plant and animal origin. Decomposing plant materials, fungi, other insects, both living and dead, all abound. Of the insects occupying this habitat, the majority are scavengers; many are fungus feeders; some are semipredators, feeding on both dead and living materials; a few are strictly predators as larvae or adults or both. Associated with stored food products are species from each of these sources. The species listed below, with the exception of the Tenebrionidae, fall largely in the last category.

Coleoptera	Coleoptera—Continued
Corynetidae	Cucujidae
<i>Corynetes coeruleus</i> Dej.	<i>Laemophloeus ferrugineus</i> (Steph.)
Ostomatidae	<i>Laemophloeus minutus</i> (Oliv.)
<i>Temnochila coerulea</i> (Oliv.)	<i>Laemophloeus ater</i> (Oliv.)
<i>Tenebroides mauritanicus</i> (Linn.)	<i>Laemophloeus janeti</i> Grouv.
<i>Tenebroides manus</i> (Melsh.)	<i>Laemophloeus turcicus</i> Grouv.
<i>Tenebroides corticalis</i> (Melsh.)	<i>Laemotmetus rhizophagoides</i> (Walk.)

## Coleoptera—Continued

## Tenebrionidae

- Uloa culinaris* (Linn.)  
*Gnathocerus cornutus* (Fab.)  
*Gnathocerus maxillosus* (Fab.)  
*Cynaesus angustus* (Lee.)  
*Latheticus oryzae* Waterh.  
*Tribolium castaneum* (Hbst.)

## Tenebrionidae—Continued

- Tribolium confusum* duV.  
*Tribolium madens* Charp.  
*Palorus ratzeburgi* Wissm.  
*Palorus subdepressus* Woll.  
*Aphanotus destructor* (Uytt.)  
*Sitophagus hololeptoides* (Cast.)  
*Platydemus ruficornis* Sturm

**Wood Borers and Wood Scavengers.**—A very few of the insects that attack stored food products were derived from wood-boring types. Most of these retain boring characteristics and attack either whole grains and seeds or processed foods of a compact nature. The ptnids and anobiids also infest loosely packed and ground products.

## Coleoptera

## Ptnidae

- ?*Ptinus* spp.

## Anobiidae

- Lasioderma serricorne* (Fab.)  
*Stegobium paniceum* (Linn.)  
*Catorama mexicana* Chev.  
*Catorama punctulata* Lee.

## Bostrichidae

- Rhyzopertha dominica* (Fab.)  
*Dinoderus bifoveolatus* Woll.  
*Dinoderus minutus* (Fab.)  
*Protostephanus truncatus* (Horn)

## Scolytidae

- Pagiocerus frontalis* (Fab.)  
*Pagiocerus rimosus* Eich.  
*Pagiocerus zeae* Eggers

**Scavengers or Depredators in the Nests of Other Insects.**—The nests of bees, wasps, ants, and certain other insects present an advantageous environment for many insects. This habitat offers protection, favorable temperatures and humidities (sometimes actually kept under some degree of control), and usually stored food materials (pollen, seeds, dead insects). Several stored-products pests may have come originally from this environment; others (dermestids, cucujids, silvanids, tenebrionids, anobiids) find it favorable.

Pollen stored in bees' nests may have greatly affected the evolution of insect food habits by bridging the gap between plant and animal foods. Pollen has a high protein content and in many of its chemical properties resembles animal materials on the one hand, and grains, seeds, and certain other concentrated plant products on the other. Many predaceous or parasitic insects can subsist wholly or partially on pollen—for example the ichneumonid genus *Grotea*, members of which are bee-egg predators in the primary larval stage, pollen feeders in later instars. The clerids as a group must be classed as predators; and the genus *Trichodes* contains many species predaceous on larvae of bees and wasps, or on eggs of grasshoppers. Yet *Trichodes* larvae, apparently unable to subsist on other plant materials, can be reared on pollen alone, though this diet greatly retards their development rate. Meloid beetles also live either as predators on grasshopper eggs or in bees' nests. As with *Grotea*, the primary larvae are egg predators, and subsequent instars are pollen feeders. Other groups of insects exhibit similar habits, a fact indicating the dietary relation between pollen and animal food (especially insect eggs). At least two groups of stored-products pests—the ptnids and the dermestids—bridge the gap between plant and animal foods; both these groups are found commonly in bees' nests, subsisting wholly or partially on pollen.

Apparently, gregarious bees (especially anthophorids), associates of gregarious bees (especially *Osmia*), social bees (*Apis*, *Bombus*), and social wasps



(*Vespa*, *Vespula*, *Dolichovespula*) provide more favorable conditions for stored-food-inhabiting insects than do other groups of bees and wasps. The explanation, undoubtedly, is that such species use their nesting sites for longer periods, develop larger and more concentrated populations, and store more food than do the more solitary forms. The following groups appear to be referable to this habitat.

Lepidoptera

Galleriidae

*Achroia grisella* (Fab.)

*Galleria mellonella* (Linn.)

Phycitidae

*Vitula serratilineella* Rag.

*Vitula edmandsii* (Pack.)

Coleoptera

Dermestidae

*Trogoderma* spp.

*Anthrenus* spp.

*Attagenus* spp.

Ptinidae

*Ptinus* spp.

*Parasites and Predators.*—Among the hundreds of parasites and predators associated with stored food products (Cotton and Good, 1937; Clausen, 1940), it is not known which species are attracted to their hosts by the food materials and which by the environment in which these are stored. The primary attraction seems clear, however, in some of the coleopterous predators (especially Histeridae, Ostomatidae, Cleridae, Corynetidae). The ostomatids and clerids are primarily bark and wood inhabitants; the histerids live in nests of birds, mammals, aculeate Hymenoptera, and so forth; and the corynetids in both types of habitat. The anthocorid bugs exhibit a similar pattern: *Lyctocoris campestris* (Fab.) occurs under bark, in birds' nests, and in nests of wasps and bumblebees.

Corbett, Yusope, and Hassan (1937) found that *Necrobia rufipes* prefers decomposed, moldy copra. Apparently, however, it is attracted not by the molds, coconut oil, or fatty acids, but by the decomposition products of the molds. Conceivably, the beetles associate these products with the presence of *Carpophilus* and other larvae upon which they are predaceous. This species and other cosmopolitan, semipredaceous *Necrobia* are probably attracted by the habitat (animal and oleaginous plant products) rather than by the insects (*Dermestes*, *Piophilus*, *Carpophilus*) which constitute much of their diet. The same is probably true of the window fly, *Scenopinus fenestralis* Linn., the larvae of which feed on mites and other small insects in various animal and plant products (hair, feathers, grain, farinaceous foodstuffs, and the like).

#### NATURAL RESERVOIRS FOR STORED-PRODUCTS INSECTS

Closely tied up with the origin and evolution of stored-food-products insects is the problem of natural reservoirs. Insufficient attention has been given to the occurrence of these insects in nature and to the role of their habitats as reservoirs that may maintain foci for future infestations. Balzer (1942) remarks: "Stored grain insects can and do enter warehouses and mills by flight. In an experiment conducted during the summer of 1937, nine species of stored grain insects, including the worst enemies of milled and rough rice, were collected in insect flight traps at two rice mills." Since these insects do migrate into storage units, natural reservoirs may perhaps be important in maintaining pest populations.

*Bark and Decomposing Wood Habitat.*—The pests of stored food products

have an important natural habitat beneath the bark of trees—a protected environment with favorable temperatures and humidities approaching those of many food-storage units. Unfortunately, records of their occurrence under bark or in wood are scanty and widely scattered; yet in the writer's experience they are not at all uncommon in this environment. Among the species recorded from under bark and for which this habitat may possibly act as a reservoir are the following:

## Thysanura

## Lepismatidae

## Blattaria

## Blattidae

## Corrodentia

## Cacciliidae

## Liposcelidae

## Atropidae

## Lepidoptera

## Galleriidae

*Aphomia sociella* (Linn.) (Forbes, 1920)

## Phycitidae

*Ephestia elutella* (Hbn.) (Forbes, 1920)

## Tineidae

*Tinea cloacella* Haw. (Escherich, 1931)

*Tinea parasitella* Hbn. (Escherich, 1931)

*Tinea corticella* Curt. (Escherich, 1931)

*Tinea quercella* H. S. (Escherich, 1931)

*Tineola bisselliella* (Hüm.) (Linsley ms.)

## Coleoptera

## Histeridae

*Dendrophilus punctatus* Hbst. (Stephens, 1839; Fowler, 1889; Blatchley, 1910)

*Carcinops* sp. (Stebbing, 1914)

*Carcinops 14-striata* Steph. (Linsley ms.)

## Corynetidae

*Corynetes coeruleus* Dej. (*auct.*)

## Dermestidae

*Trogoderma versicolor* Creutz. (Blackman and Stage, 1924, as *tarsalis*)

*Trogoderma ornatum* (Say) (Auten, 1925)

*Trogoderma sternale* Jayne (Linsley ms.)

*Trogoderma* spp. (De Leon, 1939; Linsley ms.)

*Attagenus pellio* (Linn.) (Linsley ms.)

*Attagenus piceus* (Oliv.) (Linsley ms.)

*Anthrenus verbasci* (Linn.) (Linsley ms.)

*Dermestes lardarius* Linn. (Day, 1922; Walsh, 1931; Richards, 1931a)

## Ostomatidae

*Temnochila caerulea* (Oliv.) (Clausen, 1931)

*Tenebroides corticalis* (Melsh.) (Blackman and Stage, 1924; Cotton and Good, 1937)

*Tenebroides mauritanicus* (Linn.) (Stephens, 1839; Fowler, 1891; Fall, 1901)

*Tenebroides nanus* (Melsh.) (Cotton and Good, 1937)

*Lophocateres pusillus* (Klug) (Linsley ms.)

## Nitidulidae

*Carpophilus hemipterus* (Linn.) (Stebbing, 1914)

*Carpophilus* spp. (Stebbing, 1914)

## Silvanidae

*Oryzaephilus mercator* (Fauv.) (Guillebeau, 1890)

*Oryzaephilus surinamensis* (Linn.) (Fowler, 1891; Champion, 1896)

*Cathartus quadricollis* (Guer.) (Linsley ms.)

*Ahasverus advena* (Watl.) (Lucas, 1849; Fall, 1901)

*Naustibius clavicornis* Kug. (*auct.*)

## Cucujidae

- Laemophloeus ferrugineus* (Steph.) (Fowler and Donisthorpe, 1913; Picard, 1919;  
Beare and Donisthorpe, 1922; Cotton and Good, 1937)  
*Laemophloeus ater* (Oliv.) (Perris, 1853; Fowler, 1889; Picard, 1919)

## Cryptophagidae

- Cryptophagus* spp. (duct.)

## Mycetophagidae

- Typhaea stercorea* (Linn.) (Beare and Donisthorpe, 1922)  
*Litargus balteatus* Lec. (Linsley ms.)

## Lathridiidae

- Holoparamesus caularum* (Aubé) (Fall, 1899)  
*Holoparamesus depressus* Curtis (Hinton, 1941b)  
*Coninomus constrictus* (Gyll.) (Fowler, 1889)  
*Coninomus nodifer* (Westw.) (Reitter, 1911)  
*Enicmus minutus* (Linn.) (Beare and Donisthorpe, 1922)  
*Corticaria fenestralis* (Linn.) (Donisthorpe, 1939)

## Tenebrionidae

- Alphitophagus bifasciatus* (Say) (Mulsant, 1854, *et auct.*)  
*Platydemus ruficornis* Sturm (Blatchley, 1910)  
*Doliema pallida* (Say) (Blatchley, 1910)  
*Doliema plana* (Fab.) (Fall, 1901; Ting, *in litt.*)  
*Gnathocerus cornutus* (Fab.) (Lucas, 1849; Chittenden, 1895b; Essig, 1926)  
*Gnathocerus maxillosus* (Fab.) (Wollaston, 1860; De Leon, *in litt.*)  
*Latheticus oryzae* Waterh. (Blair, 1930)  
*Latheticus prosopis* Chitt. (Chittenden, 1904)  
*Tribolium castaneum* (Hbst.) (Stephens, 1830; Lucas, 1849; Fowler, 1891;  
Fall, 1901; Stebbing, 1914; Blair, 1930)  
*Tribolium confusum* duV. (Stebbing, 1914; Van Dyke, *in litt.*; Linsley ms.)  
*Tribolium madens* Charp. (Redtenbacher, 1858; Good, 1933; De Leon, 1939)  
*Tribolium indicum* Blair (Blair, 1930)  
*Aphanotus parallelus* Csy. (Linsley ms.)  
*Aphanotus brevicornis* (Lec.) (De Leon, *in litt.*; Van Dyke, *in litt.*; Blaisdell,  
*in litt.*; Linsley ms.)  
*Aphanotus destructor* (Uytt.) (Linsley ms.)  
*Cynaesus angustus* (Lec.) (Van Dyke, *in litt.*; Blaisdell, *in litt.*; Linsley ms.)  
*Cynaesus depressus* Horn (Van Dyke, *in litt.*)  
*Palorus ratzeburgi* Wissm. (Stephens, 1832; Wissmann, 1848; Champion, 1896;  
Fowler and Donisthorpe, 1913; Swezey, 1942)  
*Palorus depressus* (Fab.) (Fowler, 1891; Champion, 1896; Blair, 1930)  
*Palorus subdepressus* Woll. (Champion, 1896)  
*Palorus* spp. (Blair, 1930)  
*Uloma culinaris* (Linn.) (Zacher, 1927)  
*Alphitobius diaperinus* (Panz.) (Fall, 1901; Linsley ms.)  
*Alphitobius piceus* (Oliv.) (Stebbing, 1914)  
*Sitophagus hololeptoides* (Cast.) (Linsley ms.)  
*Tenebrio picipes* Hbst. (Fall, 1901; Blatchley, 1910; Blackman and Stage, 1924)

## Ptinidae

- Gibbium* sp. (Stebbing, 1914)  
*Ptinus fur* (Linn.) (Fowler, 1890)  
*Ptinus palliatus* Perris (Hinton, 1941a)  
*Ptinus rufipes* Oliv. (Hinton, 1941a)  
*Ptinus lichenum* Marsh. (Picard, 1919)  
*Ptinus hirtellus* Sturm (Fowler, 1890)  
*Ptinus pusillus* Sturm (Blair, 1930)  
*Ptinus subpilosus* Sturm (Fowler, 1890)  
*Ptinus* sp. (De Leon, *in litt.*)  
*Tipnus unicolor* Pill. and Mitt. (Fowler, 1890)

## Anobiidae

- Stegobium paniceum* (Linn.) (Stebbing, 1914; Linsley, 1942c, 1942d)



## Bostrichidae

*Rhyzopertha dominica* (Fab.) (Gorham, 1883; Lesne, 1924; Froggatt, 1918)*Dinoderus* spp. (auct.)*Protostephanus truncatus* (Horn) (auct.)

*Insect Nests and Food Caches*.—Bee, wasp, and ant nests provide a favorable environment for other insects. Moderate temperatures, favorable humidities, and stores of food materials (pollen, seeds, dead insects) offer conditions not unlike those encountered in warehouses and human habitations. As a result, stored-products pests (and certain other household insects) are not uncommon in such nests. Unfortunately, most observers either have failed to record their occurrence or have noted them only in passing, except in the hive of the honeybee, where the presence of pests has some economic significance. Even in apicultural literature published data are inadequate, the more typical pests of stored products having been commonly confused with certain other inhabitants of the hive, especially wax moths.

In the spring of 1941, in connection with another study, eight nesting sites of *Anthophora* (five species) in seven localities in central and southern California were examined. Dermestids of several types abounded in all the sites; the California spider beetle, *Ptinus californicus*, and the confused flour beetle, *Tribolium confusum*, were present in three; the Indian-meal moth, *Plodia interpunctella*, the drugstore beetle, *Stegobium paniceum*, and the false black flour beetle, *Aphanotus destructor* in two; and the saw-toothed grain beetle, *Oryzaephilus surinamensis*, and the webbing clothes moth, *Tincola bisselliella*, each in one site. In a single colony on the edge of the Mojave Desert about 10 miles from the nearest human habitation were *S. paniceum*, *O. surinamensis*, and one of the dermestid beetles, *Trogoderma ajax*; and in a second area, only slightly less isolated, *Plodia interpunctella* and *Tincola bisselliella*.

Of these insects, *Ptinus californicus* is a native species which seems to be definitely associated with bees, never having been recorded as breeding outside of bees' nests. It is most common as a pollen feeder in cells of *Osmia*, especially where nesting sites are used repeatedly year after year. In the laboratory, however, it can be readily reared on cornmeal, whole wheat flour, and other cereal products. *Trogoderma ajax* is also a native species. It is the commonest dermestid associated with anthophorid bees in California and has been found infesting grain. *Aphanotus destructor* is a common pest of many stored products, especially dog biscuits, rabbit food, and various cereal products. Of the remaining species, *Plodia interpunctella*, *Stegobium paniceum*, and *Oryzaephilus surinamensis* live on a wide variety of plant products, apparently finding the pollen stored by bees one of many suitable foods. *Tincola bisselliella* lives as a scavenger on products of animal origin. Probably no one of these last species is native to California; yet their presence even in isolated habitats suggests that they find the environment of bees' nests satisfactory and may possibly have occupied it for a long period of time.

Among the stored-products insects and their relatives that breed or occur in nests of ants, bees, and wasps, the following may be noted:

## Acarina

## Acaridae

*Tyrophagus* sp. (bees, Linsley ms.)

## Thysanura

## Lepismatidae

*Lepisma saccharina* Linn. (*Vespa*, Tuck, 1896; *Anthophora*, *Osmia*, Linsley ms.)

*Ctenolepisma urbani* Slaugh. (*Anthophora*, Linsley ms.)

## Corrodentia

## Atropidae

*Atropos pulsatorium* (Linn.) (*Apis*, *Polistes*, Essig, 1942)

## Liposcelidae

*Liposcelis divinatorius* (Müll.) (bees, Linsley, 1942a)

*Liposcelis formicarius* (Hagen) (ants, Essig, 1942)

## Lepidoptera

## Phycitidae

*Plodia interpunctella* (Hbn.) (*Bombus*, Frison, 1926; *Apis*, auct.; *Anthophora* and *Osmia*, Linsley and MacSwain, 1942b)

*Ephestia kühniella* Zell. (*Bombus*, Davidson, 1894; *Apis*, auct.; *Vespa* and *Vespula*, auct.)

*Ephestia cautella* (Walk.) (*Apis*, Milum, 1940a)

*Ephestia elutella* (Hbn.) (*Apis*, Laboulbene, 1864)

*Vitula serratilineella* Rag. (*Apis*, Cockle, 1920; *Xylocopa*, Linsley, 1943)

*Vitula edmandsii* (Pack.) (*Bombus*, Packard, 1865, et auct.; *Apis*, Milum, 1940b)

## Galleriidae

*Achroia grisella* (Fab.) (*Apis*, auct.)

*Aphomia sociella* (Linn.) (*Bombus*, *Apis*, *Vespa*, auct.)

## Pyraustidae

*Chalcoela iphitalis* (Walk.) (bees, Linsley, 1942a)

*Dicymolomia pegasalis* (Walk.) (bees, Linsley, 1942a)

## Oecophoridae

*Hofmannophila pseudospretella* (Staint.) (*Apis*, Hinton, 1943a)

*Endrosis lacteella* (D. and S.) (*Acanthomyops*, Donisthorpe, 1927b; *Sceliphron*, Linsley ms.)

## Tineidae

*Tineola bisselliella* (Hüm.) (*Formica*, Donisthorpe, 1927b; *Anthophora*, Linsley and MacSwain, 1942b)

*Trichophaga tapetzella* (Linn.) (*Sceliphron*, Linsley ms.)

## Coleoptera

## Histeridae

*Dendrophilus punctatus* Hbst. (*Formica*, Fowler, 1889; *Lasius*, Donisthorpe, 1927b)

*Dendrophilus pygmaeus* (Linn.) (*Formica*, Fowler, 1889; Donisthorpe, 1927b)

## Corynetidae

*Corynetes coeruleus* (Dej.) (*Vespa*, Butler, 1896)

## Dermestidae

*Attagenus pelli* (Linn.) (*Apis*, Hinton, 1943a)

*Attagenus pantherinus* Ahrens (bees, wasps, Hinton, in litt.)

*Attagenus piceus* (Oliv.) (*Anthophora*, *Osmia*, Linsley and MacSwain, 1942b)

*Megatoma undata* (Linn.) (*Andrena*, Fowler and Donisthorpe, 1913)

*Perimegatoma vespulae* Milliron (*Dolichovespula*, Milliron, 1939)

*Perimegatoma variegatum* Horn (*Sceliphron*, Linsley ms.)

*Perimegatoma jaynei* Csy. (*Sceliphron*, Linsley ms.)

*Trogoderma glabrum* Hbst. (*Osmia*, *Odynerus*, Maréchal, 1932)

*Trogoderma ornatum* (Say) (*Apis*, Chittenden, 1897; Patton, 1931)

*Trogoderma versicolor* Creutz. (*Pelopoeus*, *Odynerus*, *Anthophora*, etc., Riley, 1883; *Sceliphron*, Linsley ms.)

*Trogoderma ajax* Csy. (*Anthophora*, Linsley and MacSwain, 1942b)

*Trogoderma sternale* Jayne (*Dolichovespula*, *Sceliphron*, *Osmia*, Linsley ms.)

*Trogoderma socium* Lea (*Iridomyrmex*, Lea, 1905, 1911)

*Anthrenus muscorum* (Linn.) (bees, wasps, Hinton, in litt.)

*Anthrenus fuscus* (Latr.) (*Osmia*, Maréchal, 1932)

*Anthrenus verbasci* (Linn.) (*Osmia*, Linsley and MacSwain, 1942a; *Polistes*, *Dolichovespula*, *Sceliphron*, Linsley ms.)

- Anthrenus* spp. (*Anthophora*, Hicks, 1926; Mickel, 1928)  
*Dermestes lardarius* Linn. (*Apis*, Riley, 1870; Lintner, 1890; Hinton, 1943a)  
*Dermestes murinus* Linn. (*Philanthus*, Latreille, 1802)  
*Globicornis corticalis* (Eich.) (bees, wasps, Hinton, *in litt.*)

## Nitidulidae

- Brachypeplus inquilinus* Lea (*Apis*, Lea, 1912)  
*Brachypeplus auritus* Mur. (*Trigona*, Lea, 1911)  
*Brachypeplus basalis* Er. (*Trigona*, Lea, 1911)  
*Brachypeplus blandus* Mur. (*Apis*, Lea, 1912)  
*Aethina tumida* Mur. (*Apis*, Lundie, 1940)  
*Carpophilus planatus* Mur. (*Trigona*, Lea, 1912)

## Silvanidae

- Oryzaephilus surinamensis* (Linn.) (*Anthophora*, Linsley and MacSwain, 1942b)

## Cucujidae

- Laemophloeus ferrugineus* (Steph.) (*Vespula*, Tuck, 1896)  
*Cryptamorpha delicata* Blackb. (*Ectatomma*, Lea, 1912)

## Cryptophagidae

- Cryptophagus setulosus* Sturm (*Vespula*, Tuck, 1896; *Bombus*, Tuck, 1896; Sladen, 1912)  
*Cryptophagus populi* Payk. (*Colletes* and *Dasypoda*, Champion, 1918)  
*Cryptophagus laevendali* Ganglb. (*Vespa*, Donisthorpe, 1927a)  
*Cryptophagus pubescens* Sturm (*Vespula*, Tuck, 1896; Britten, 1911; Donisthorpe, 1927a)  
*Cryptophagus umbratus* Er. (*Bombus*, Tuck, 1896)  
*Cryptophagus punctipennis* Bris. (*Bombus*, Tuck, 1896)  
*Cryptophagus pilosus* Gyll. (*Vespula*, Tuck, 1896)  
*Cryptophagus scanicus* (Linn.) (*Vespula*, *Bombus*, Tuck, 1896; *Acanthomyops*, Walker, 1920)  
*Cryptophagus saginatus* Sturm (*Bombus*, Tuck, 1896)  
*Cryptophagus distinguendus* Sturm (*Vespula*, Tuck, 1897; *Bombus*, Fowler and Donisthorpe, 1913)  
*Cryptophagus badius* Sturm (*Vespa*, Tuck, 1897)  
*Cryptophagus dentatus* Hbst. (*Acanthomyops*, Walker, 1920b)  
*Cryptophagus affinis* Sturm (*Acanthomyops*, Walker, 1920b)  
*Cryptophagus fumatus* Sturm (*Acanthomyops*, Walker, 1920b)  
*Cryptophagus croceus* Zimm (*Bombus*, Frison, 1926)  
*Cryptophagus cellaris* (Scop.) (*Apis*, Falcoz, 1929-1931)  
*Atomaria ruficornis* (Marsh.) (*Vespula*, Tuck, 1896)  
*Atomaria peltata* Kraatz (*Vespula*, Tuck, 1896)  
*Antherophagus pallens* (Fab.) (*Bombus*, Tuck, 1896; Sladen, 1912 [as *silaceus*])  
*Antherophagus nigricornis* (Fab.) (*Bombus*, Tuck, 1896; Sladen, 1912)  
*Antherophagus ochraceus* Melsb. (*Bombus*, Packard, 1865; Frison, 1926; Plath, 1934)

## Mycetophagidae

- Mycetophagus quadriguttatus* Müll. (*Vespa*, Tuck, 1896; *Acanthomyops*, Walker, 1920b)

## Lathridiidae

- Coninomus constrictus* (Gyll.) (*Acanthomyops*, Walker, 1920b)  
*Coninomus nodifer* (Westw.) (*Bombus*, Frison, 1926)  
*Enicmus minutus* (Linn.) (*Bombus*, *Vespa*, Morley, 1899; *Acanthomyops*, Walker, 1920b)  
*Enicmus transversus* Oliv. (*Acanthomyops*, Walker, 1920b)  
*Enicmus histrio* Joy (*Acanthomyops*, Walker, 1920b)

## Tenebrionidae

- Tenebrio molitor* Linn. (*Apis*, Hinton, 1943a)  
*Tenebrio obscurus* Fab. (*Bombus*, Frison, 1926)  
*Tenebrio tenebroides* Beauv. (*Bombus*, St. George, 1925; Frison, 1926)  
*Tribolium castaneum* (Hbst.) (*Megachile*, Kotinsky, 1907)  
*Tribolium confusum* duRoi. (*Clisodon*, *Anthophora*, *Osmia*, Linsley and MacSwain, 1942a)



*Tribolium madens* Charp. (*Apis*, Leech, 1943)

*Tribolium myrmecophilum* Lea (*Iridomyrmex*, Lea, 1905)

*Aphanotus brevicornis* (Lec.) (*Xylocopa*, Davidson, 1893; Nininger, 1916; Linsley, 1943; *Anthidium*, Hicks, 1929)

*Aphanotus destructor* (Uytt.) (*Osmia*, *Anthophora*, Linsley, 1942a)

*Cynaesus augustus* (Lec.) (*Osmia*, Linsley, 1942a)

#### Ptinidae

*Ptinus fur* (Linn.) (*Osmia*, Enslin, 1925; Schmitt, 1929; *Chelostoma*, Maréchal, 1932; *Vespula*, Tuck, 1896; *Vespa*, Tuck, 1896; Butler, 1896; and Husing, 1935; *Apis*, Hinton, 1943a)

*Ptinus sexpunctatus* Panz. (*Megachile*, Bedel, 1872; Redtenbacher, 1874; Maneval, 1929; *Osmia*, Hamm, 1924; Maneval, 1929; Maréchal, 1932; *Hoplitis*, Maneval, 1929; *Chelostoma*, Maneval, 1929; *Bombus*, Fowler, 1889; *Lasius*, Walker, 1920b) *Ptinus subpilosus* Sturm (*Acanthomyops*, Fowler, 1890; Walker, 1920b; Donisthorpe, 1927b)

*Ptinus hirtellus* Sturm (*Sceliphron*, Linsley ms.)

*Ptinus vaulogerii* Pic (*Osmia*, Cros, 1932)

*Ptinus californicus* Pic (*Osmia*, *Anthophora*, Linsley and MacSwain, 1942a)

*Ptinus raptor* (Linn.) (*Apis*, Arnhart, 1929; Brassler, 1929)

*Ptinus exulans* Er. (*Apis*, Rayment, 1935)

*Niptus hololeucus* (Fald.) (*Dolichovespula*, Tuck, 1897)

*Diphobia familiaris* Oll. (ants, Lea, 1905)

*Pausoptinus laticornis* Lea (*Iridomyrmex*, *Cremastogaster*, Lea, 1905)

*Polyplocotes longicollis* Westw. (ants, Lea, 1911)

*Polyplocotes nitidus* Westw. (ants, Lea, 1911)

*Diplocotes howittanus* Westw. (ants, Lea, 1911)

*Diplocotes foveicollis* Oll. (ants, Lea, 1911)

*Diplocotes armicollis* Lea (ants, Lea, 1911)

*Ectrephes formicarium* Pasc. (ants, Lea, 1911)

*Ectrephes kingii* Westw. (ants, Lea, 1911)

*Hexaplocotes sulcifrons* Lea (ants, Lea, 1911)

#### Anobiidae

*Stegobium paniceum* (Linn.) (*Anthophora*, *Osmia*, Linsley and MacSwain, 1942b)

*Nests and Food Caches of Birds and Mammals*.—Undoubtedly a great many household pests live in the nests of birds and mammals. Included are many insect inhabitants of stored food products. A few are listed below.

#### Thysanura

##### Lepismatidae

*Sp. indet.* (woodrat, Davis, 1934)

#### Corrodentia

##### Liposcelidae

*Sp. indet.* (woodrat, Davis, 1934)

*Liposcelis divinatorius* (Müll.) (purple martin, bluebird, house wren, crested flycatcher, McAtee, 1927; starling, English sparrow, McAtee, 1929)

#### Coleoptera

##### Histeridae

*Dendrophilus punctatus* Hbst. (owl, Walker, 1896, 1904; Bickhardt, 1907; woodpecker, Auzat, 1925; Ross, 1940; jackdaw, hawk, screech owl, pigeon, Auzat, 1925; crow, hoopoe, Bickhardt, 1916; starling, Joy, 1905; Auzat, 1925; McAtee, 1929; squirrel, Donisthorpe, 1927b)

*Dendrophilus opacus* Ross (woodrat, Ross, 1940)

*Dendrophilus tularensis* Ross (ground squirrel, Wenzel, *in litt.*)

*Saprinus semistriatus* Scriba (prairie dog, Brown, 1928)

*Carcinops 14-striata* Steph. (owl, Walker, 1896, Bickhardt, 1907; starling, purple martin, McAtee, 1929)

##### Dermestidae

*Trogoderma ornatum* (Say) (English sparrow, house wren, McAtee, 1929)

- Trogoderma versicolor* Creutz. (English sparrow, house wren, McAtee, 1929)  
*Trogoderma sternale* Jayne (Cliff swallow, Linsley ms.)  
*Trogoderma* sp. (purple martin, McAtee, 1929)  
*Anthrenus verbasci* (Linn.) (sparrow, Yokoyama, 1929; Herfs, 1936)  
*Anthrenus fuscus* (Latr.) (sparrow, Yokoyama, 1929; Herfs, 1936)  
*Anthrenus serophulariae* (Linn.) (sparrow, Herfs, 1936)  
*Anthrenus pimpinellae* (Fab.) (sparrow, Yokoyama, 1929; Herfs, 1936)  
*Anthrenus occidentis* Csy. (magpie, Jellison and Philip, 1933; English sparrow, house finch, black phoebe, barn swallow, Linsley ms.)  
*Attagenus piceus* (Oliv.) (chipmunk, Linsley, 1942b; birds, Back and Cotton, 1938; starling, English sparrow, purple martin, house wren, bluebird, McAtee, 1929)  
*Attagenus schäfferi* Hbst. (chipmunk, English sparrow, house finch, Linsley ms.)  
*Attagenus pellio* (Linn.) (birds, Zacher, 1939)  
*Attagenus elongatulus* Csy. (woodrat, Davis, 1934)  
*Attagenus nigripes* Casey (English sparrow, house finch, black phoebe, cliff swallow, Linsley ms.)  
*Attagenus japonicus* Reitt. (sparrow, Yokoyama, 1929; Herfs, 1936)  
*Dermestes elongatus* Lee. (black-crowned night heron, Barber, 1914)  
*Dermestes signatus* Lec. (crow and magpie, Jellison and Philip, 1933; house finch, Linsley ms.)  
*Dermestes talpinus* Mann. (magpie, Jellison and Philip, 1933)  
*Dermestes bicolor* Fab. (pigeon loft, Bickhardt, 1907)

## Lathridiidae

- Metophtalmus rudis* Fall (woodrat, Davis, 1934)  
*Metophtalmus trux* Fall (woodrat, Davis, 1934)  
*Coninomus nodifer* Westw. (mole, Bickhardt, 1907; woodrat, Davis, 1934)  
*Melanophthalma villosa* Zimm. (woodrat, Davis, 1934)  
*Melanophthalma americana* Mann. (woodrat, Davis, 1934)  
*Melanophthalma distinguenda* Com. (woodrat, Davis, 1934; Linsley ms.)  
*Melanophthalma similata* Gyll. (woodrat, Davis, 1934; Linsley ms.)  
*Melanophthalma* sp. (robin, Dobrosky, 1925)  
*Enicmus minutus* (Linn.) (stork, Schaufuss, 1916)  
*Enicmus crenatus* Lec. (woodrat, Davis, 1934)  
*Enicmus suspectus* Fall (woodrat, Davis, 1934)  
*Cartodere ruficollis* (Marsh.) (stork, Reitter, 1911)  
*Corticaria pubescens* (Gyll.) (birds, Donisthorpe, 1939)  
*Fuchsina occulta* Fall (woodrat, Linsley ms.)

## Tenebrionidae

- Tenebrio molitor* Linn. (pigeon loft, auct.)

## Mycetaeidae

- Mycetaea hirta* Marsh. (owls, Walker, 1896)

## Cryptophagidae

- Atomaria apicalis* Er. (field mouse, Beare, 1921)  
*Atomaria fuscata* Seh. (field mouse, Beare, 1921)  
*Atomaria* sp. (woodrat, Davis, 1934)  
*Atomaria melas* Csy. (woodrat, Linsley ms.)  
*Atomaria subdentata* Csy. (woodrat, Linsley ms.)  
*Atomaria dispersa* Csy. (woodrat, Linsley ms.)  
*Anchicera fallax* Csy. (woodrat, Linsley ms.)  
*Anchicera nanula* Csy. (woodrat, Davis, 1934)  
*Cryptophagus lyraticollis* Csy. (woodrat, Linsley ms.)  
*Cryptophagus badius* Sturm (owl, Fowler and Donisthorpe, 1913)  
*Cryptophagus umbratus* Er. (rabbit, Bickhardt, 1907)  
*Cryptophagus schmidtii* Sturm (marmot, Bickhardt, 1907)  
*Cryptophagus* sp. (woodrat, Davis, 1934)  
*Cryptophagus setulosus* Sturm (field mouse, Beare, 1921)  
*Cryptophagus pubescens* Sturm (field mouse, Beare, 1921)

## Ptinidae

- Ptinus agnatus* Fall (woodrat, Davis, 1934)

- Ptinus hirtellus* Sturm (rats, Pilate, 1905; English sparrow, McAtee, 1929)  
*Ptinus fur* (Linn.) (rats, Pilate, 1905; robin, Dobroscky, 1925; house wren, bluebird, McAtee, 1929)  
*Ptinus gandolphei* Pic (barn owl, Linsley ms.)  
*Ptinus sexpunctatus* Panz. (house martin, Blair, 1924)  
*Tipnus unicolor* Pill and Mitt. (birds, Fowler, 1890; house martin, Hinton, 1941a)  
*Mezium americanum* Lap. (sparrow, Zimmerman, 1940)  
*Mezium affine* Boield (rats, Linsley ms.)

#### Curculionidae

- Sitophilus granarius* (Linn.) (ground squirrel, Davis and Sloop, 1934)

#### Lepidoptera

##### Tineidae

- Tinea fuscipunctella* Haw. (starling, English sparrow, purple martin, house wren, bluebird, crested flycatcher, McAtee, 1929)  
*Tineola bisselliella* (Hüm.) (kangaroo rat, Hubbard, 1899)

### SUMMARY

Stored-food-products insects are widely distributed in nature as well as in the warehouses, granaries, and households of mankind. This fact is probably due to a broad range of tolerance for physical factors of the environment, a wide range of food habits, and the optimum conditions for existence frequently provided by these man-made structures.

Several of the species involved (*Bruchidae*, *Sitophilus*, spp., *Sitotroga*, spp., and so forth) infest seeds in nature. These insects have become stored-seed pests without having to undergo further evolution of food habits.

Many insects (*Cryptophagidae*, *Mycetophagidae*, *Lathridiidae*, *Acaridae*) associated with stored products are actually fungus feeders and seek out the molds and fungi existing in the foodstuffs. In nature these species are found under bark; in nests of birds, mammals, and insects; in decomposing plant material; or in any other places where molds and fungi are present.

Many food-products pests are general scavengers on dead plant materials, principally moths (*Phycitids*, *Pyralids*), but also a few other groups (*Anobiids*, *Nitidulids*).

Scavengers on dead animal materials provide a similar source of pests. Included are *Dermestes*, *Piophilus*, *Tineola*, and others.

An important natural source of stored-products pests lies among the scavengers or semipredators living under bark. To this source we must credit almost all the *Tenebrionidae* (flour beetles, mealworms), *Cucujidae*, and *Ostomatidae*.

The *Ptinidae*, *Anobiidae*, *Bostrichidae*, and *Scolytidae* probably had a wood-boring origin.

Insect nests also contribute a number of pests—in particular, several of the *Galleriidae*, *Phycitidae*, *Ptinidae*, and *Dermestidae*. The pollen stored by bees may sometimes have influenced the evolution of food habits by serving as a bridge between zoophagy and phytophagy.

Many insects in stored food products are predators and parasites of other insects present. A few of these, especially *Histeridae*, *Corynetidae*, *Anthorcoridae*, are probably attracted to the products and habitat directly rather than by any specific insects present.

Primary natural reservoirs are to be found in the following habitats: (1) bark and decomposing wood; (2) nests and food caches of other insects, especially bees, wasps, and ants; and (3) nests of birds and rodents.



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